

# Complementing our immune system: the antimicrobial mode of action of nitric oxide and its potential in wound care

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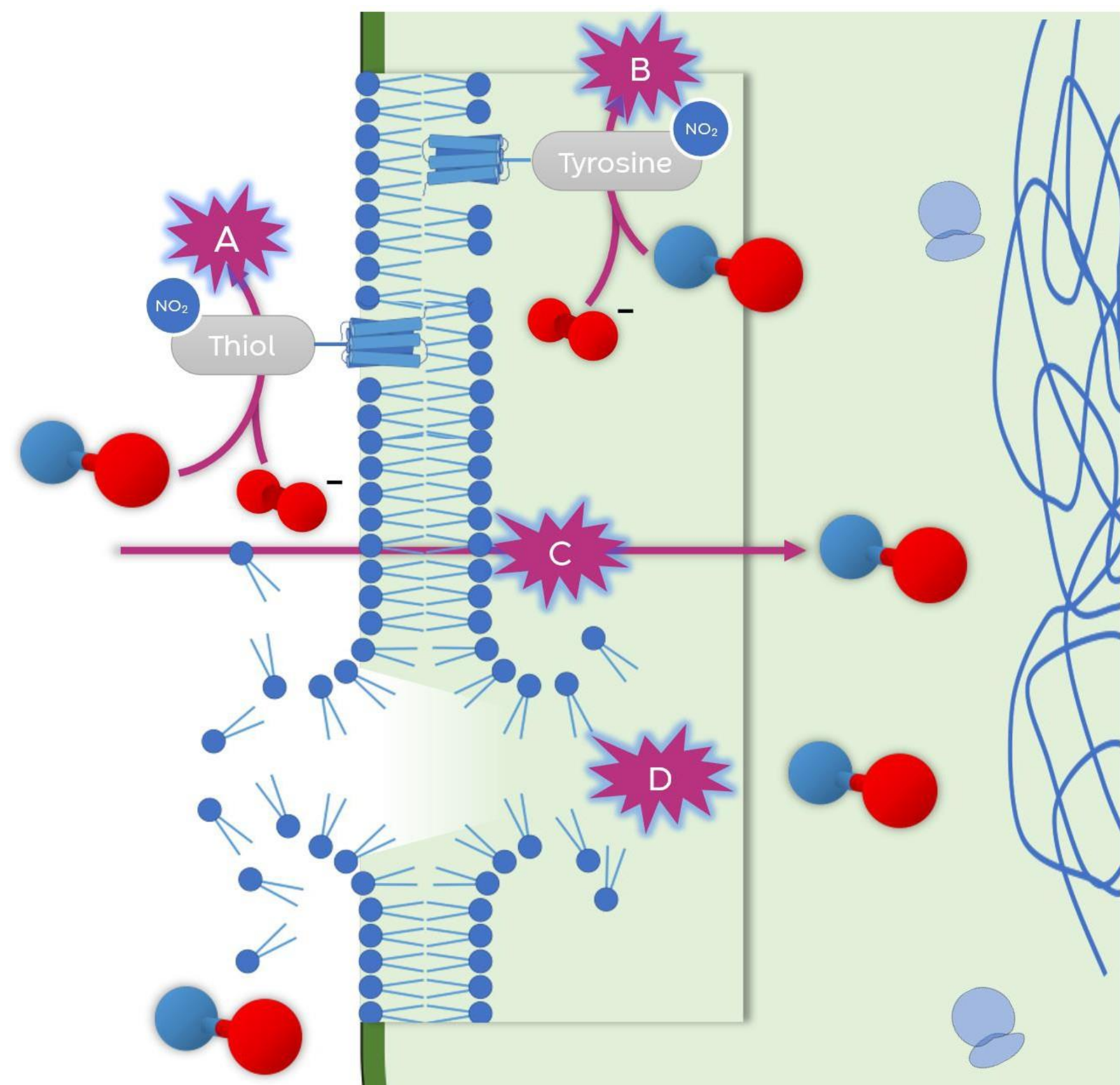
## Introduction

Nitric oxide (NO) is an innate molecule of the human immune response to invading pathogens. NO is produced in the body by nitrogen synthases (NOS) from L-arginine through a series of oxidation reactions.<sup>1</sup> Whilst NO has potent antimicrobial activity against bacteria, its effects on mammalian cells are lessened due to diffusion down the concentration gradient out of cells and into red blood cells. Here it is broken down into nitrate via oxyhemoglobin faster than it can react with intracellular components.<sup>2</sup> NO could be employed in hard-to-heal wounds that are infected or at risk of infection due to the lack of bacterial resistance mechanisms to NO and tolerance by human cells. We conducted a narrative review of the evidence underlying the mechanisms of action of NO as an antimicrobial agent in wound dressings for the treatment of hard-to-heal wounds, such as diabetic foot ulcers (DFUs)

Aim: To evaluate the evidence for nitric oxide (NO) as an antimicrobial agent for the treatment of hard-to-heal wounds

## Interactions at the cell membrane

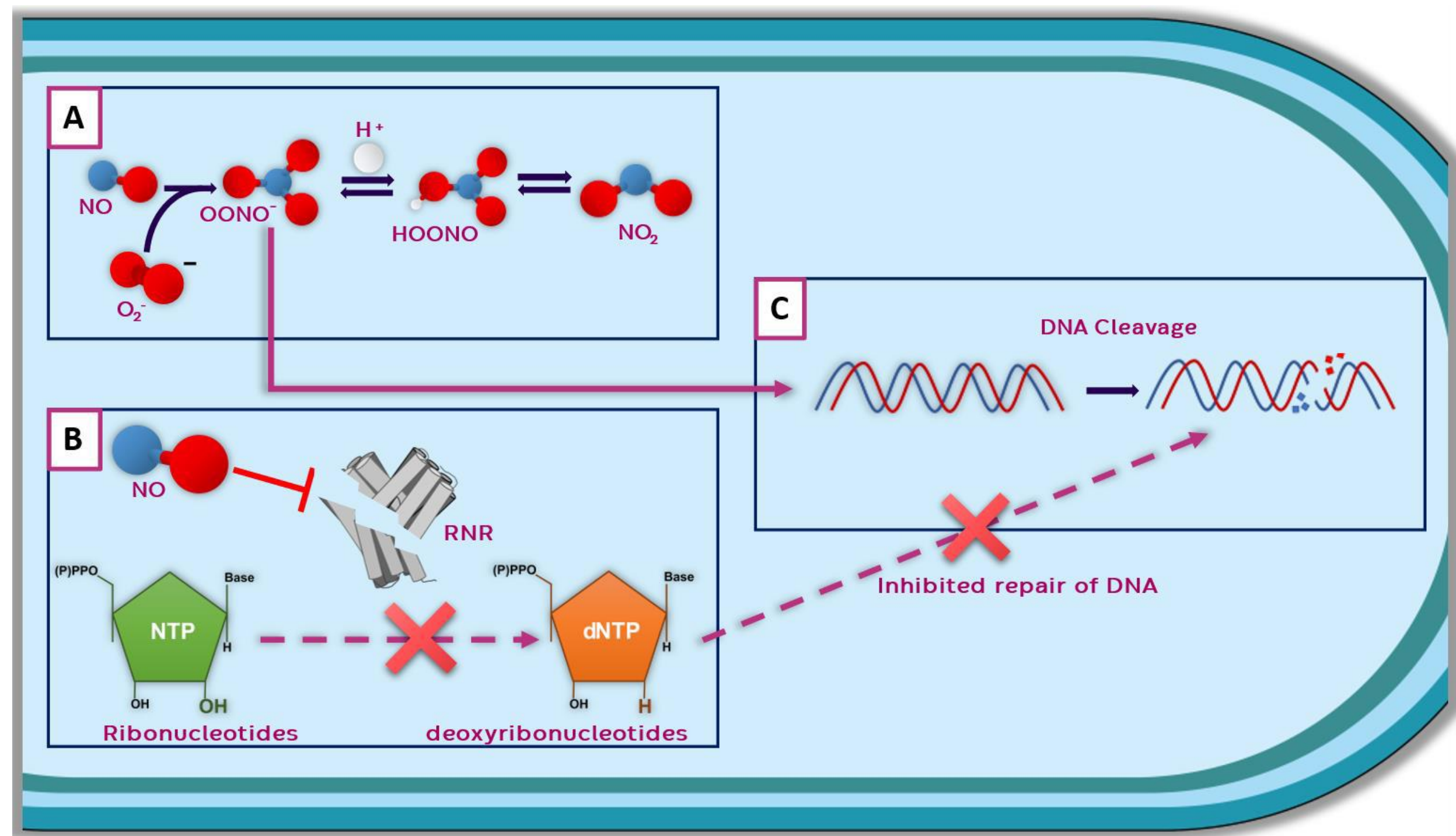
- Thiol and tyrosine group disruption (Figure 1A and 1B)**
- Reactive nitrogen species (RNS), such as dinitrogen trioxide (N<sub>2</sub>O<sub>3</sub>) and peroxynitrite (OONO<sup>-</sup>), are formed from a reaction between NO and superoxide (O<sub>2</sub><sup>-</sup>).<sup>2,3</sup>
  - N<sub>2</sub>O<sub>3</sub> has a strong affinity for thiol groups in proteins embedded within the cell membrane lipid bilayer,<sup>3</sup> adding nitrous groups, resulting in misshapen proteins, which eventually destroys their function<sup>4</sup>
  - OONO<sup>-</sup> will also react with tyrosine groups on internal-bound proteins. The end-product, 3-nitrotyrosine, is an established biomarker of cellular nitro-oxidative stress<sup>3,4</sup>
- Loss of function will result in microbial death
- Diffusion of NO through the cell wall and membrane (Figure 1C)**
- NO gas freely diffuses through bacterial cell walls and membranes<sup>5</sup> due to its small diameter, low polarity, and lipophilic property.<sup>6</sup> The rate of diffusion is affected by whether the bacteria has a thick peptidoglycan layer on the outer cell wall (Gram positive), as a thicker layer slows the rate of diffusion<sup>7</sup>
- NO gas can effectively penetrate microbial cells
- Eventual breakdown of the cell membrane (Figure 1D)**
- The tightly spaced lipid bilayer that makes up the cell membrane acts as a barrier to large and charged molecules, preventing them from entering the microbial cell. As NO disrupts both internal and external bound proteins, the cell membrane begins to break down
- The disruption of the cell membrane increases its permeability to antibiotics and other large antimicrobial molecules<sup>8</sup>



**Figure 1.** Interactions of NO and RNS formed from the reaction of NO with O<sub>2</sub> at the cell membrane. (A) Thiol group disruption. (B) Tyrosine group disruption. (C) Diffusion of NO through the cell membrane. (D) Eventual breakdown of the cell membrane

## Disruption of DNA processes

- Production of peroxynitrite (OONO<sup>-</sup>) (Figure 2A)**
- As NO moves into the microbial cell, it will form other RNS such as OONO<sup>-</sup>. RNS react with DNA, causing oxidative damage and loss of amine groups
  - OONO<sup>-</sup> has been shown to cause DNA strand breaks more efficiently than NO alone, as it reacts directly with the sugar backbone of DNA<sup>9</sup>
- Breakdown of the DNA backbone will eventually lead to DNA cleavage and an instability of the genome<sup>3,4</sup> (Figure 2C)
- Inhibition of ribonucleotide reductase (Figure 2B)**
- Ribonucleotide reductase (RNR) is an enzyme for the conversion of ribonucleotides (NTP; the building blocks of RNA), into deoxyribonucleotides (dNTP; the building blocks of DNA). dNTP in bacterial cells is essential for DNA repair and replication and overall stability of the genome<sup>10</sup>
  - NO has strong reactivity towards iron containing amino groups in the RNR enzyme, which results in its inhibition<sup>10</sup>
- Without the ability to repair and create new DNA strands, microorganisms will become unstable, unable to reproduce, and ultimately die<sup>10,11</sup>



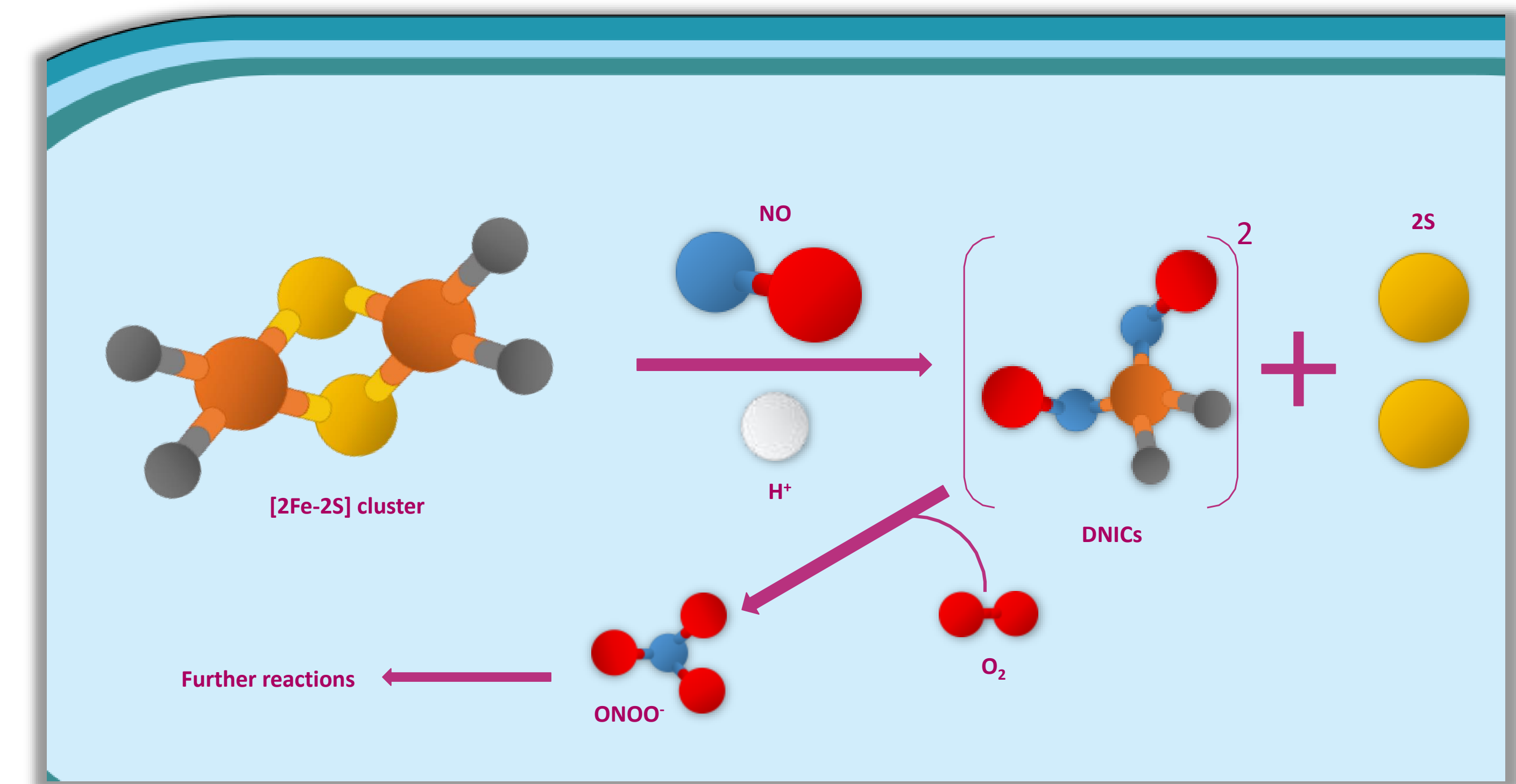
**Figure 2.** Disruption of DNA processes. (A) The production of the RNS, OONO<sup>-</sup>, from NO and O<sub>2</sub><sup>-</sup>. (B) Inhibition of RNR. (C) Cleavage of DNA by OONO<sup>-</sup> and inhibition of repair

## Discussion

The diverse microbial targets of NO and other RNS means there is less opportunity for evasion of their action, decreasing risk of resistance or reduced susceptibility. The exploitation of NO generation from exogenous sources represents a promising strategy as an antimicrobial agent, which has the potential to be used in wound dressings for the treatment of hard-to-heal wounds

## Breakdown of iron-sulfur clusters

- Iron-sulfur (Fe-S)-containing proteins are one of the primary targets of NO. Fe-S clusters are essential enzyme co-factors required for fundamental biological processes within bacteria such as oxidative phosphorylation and ribosome assembly.<sup>12</sup> Bacteria also use Fe-S clusters for electron transport, which is susceptible to NO attack<sup>13</sup>
  - NO is attracted to and reacts with sulfur, breaking the molecule into two smaller iron complexes called dinitrosyl iron complexes (DNICs), which are biomarkers for NO toxicity<sup>12</sup> (Figure 3)
  - DNICs then donate nitrogen and react with oxygen to form additional RNS species<sup>14</sup>. DNICs negatively affect bacteria cells through the release of RNS<sup>15</sup>
  - Fe released from Fe-S clusters binds to microbial DNA which is then oxidized by peroxide leading to DNA cleavage<sup>13</sup>
- Destruction of key Fe-S cluster-containing proteins disrupts pathways needed for the survival and proliferation of microorganisms



**Figure 3.** The breakdown of iron-sulfur (2Fe-2S) clusters by NO into dinitrosyl iron complexes (DNICs) and further reduction into OONO<sup>-</sup>

## References

- MacMicking et al. Nitric oxide and macrophage function. *Ann Rev Immunol* 1997; 15(1): 323-350.
- Beckman & Koppenol. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and the ugly. *Am J Physiol-Cell Physiol* 1996; 271(5): C1424-C1437.
- Carpenter & Schoenfish. Nitric oxide release: Part II. Therapeutic applications. *Chem Soc Rev* 2012; 41(10): 3742.
- Rong et al. Nitric oxide-releasing polymeric materials for antimicrobial applications: A review. *Antioxidants* 2019; 8(11): 556.
- Moller & Denicola. Diffusion of nitric oxide and oxygen in lipoproteins and membranes studied by pyrene fluorescence quenching. *Free Rad Biol Med* 2018; 128: 137-143.
- Fang. Mechanisms of nitric oxide-related antimicrobial activity. *J Clin Invest* 1997; 99(12): 2818-2825.
- Hall et al. Mode of Nitric Oxide Delivery Affects Antibacterial Action. *ACS Biomater Sci Eng* 2020; 6(1): 433-441.
- Rouillard et al. Exogenous Nitric Oxide Improves Antibiotic Susceptibility in Resistant Bacteria. *ACS Infect Dis* 2021; 7(1): 23-33.
- Burney et al. The chemistry of DNA damage from nitric oxide and peroxynitrite. *Mutat Res Mol Mech Mutagen* 1999; 424: 37-49.
- Lepoivre et al. Inactivation of ribonucleotide reductase by nitric oxide. *Biochem Biophys Res Com* 1991; 179(1): 442-448.
- Torrents. Ribonucleotide reductases: Essential enzymes for bacterial life. *Front Cell Infect Microbiol* 2014; 4: 52.
- Fitzpatrick & Kim. Synthetic Modeling Chemistry of Iron-Sulfur Clusters in Nitric Oxide Signaling. *Acc Chem Res* 2015; 48(8): 2453-2461.
- Wink et al. Nitric oxide and redox mechanisms in the immune response. *J Leukocyte Biol* 2011; 89(6): 873-891.
- Radi. Protein Tyrosine Nitration: Biochemical Mechanisms and Structural Basis of Functional Effects. *Acc Chem Res* 2013; 46(2): 550-559.
- Vanin. Physico-chemistry of dinitrosyl iron complexes as a determinant of their biological activity. *Int J Mol Sci* 2021; 22(19): 10356.