

## Abstract

**Introduction:** Bacterial resistance to antibiotics is an important problem, especially in non-healing wounds. Two of the most commons pathogens associated with these infections are *Staphylococcus aureus* and *Acinetobacter baumannii*. Exosomes are lipid bilayer-delimited particles that are naturally released from almost all types of cells but, unlike a cell, cannot replicate. They serve as a fundamental intercellular communication system and have shown efficacy in tissue repair. In this *in vitro* study, we examined the use of antibiotic-loaded exosomes formulated with synthetic liposomes to treat and prevent wound infections.

**Methods:** Fresh cultures of bacterial pathogenic isolate obtained directly from American Type Culture Collection (ATCC), Rockville, Maryland, were used in these studies, specifically Methicillin Resistant *Staphylococcus aureus* MRSA USA300 (MRSA USA300) and *Acinetobacter baumannii* ATCC 19606 (AB19606). Prior to evaluating zones of inhibition, minimal inhibitory concentrations (MIC) for each pathogen were determined. A modified Kirby Bauer technique was then used to demonstrate the efficacy of topical exosome-based formulations containing the antibiotic Moxifloxacin (MOX). Inhibition zones were measured using ImageJ. Results were tabulated, and statistical analysis was performed to demonstrate the differences between treatments.

**Results:** MIC testing resulted in an inhibitory MOX concentration of 32 µg/ml and 256 µg/ml for AB19606 and MRSA USA300, respectively. MOX alone and in combination with topical exosome-loaded gels resulted in significant ( $p \leq 0.05$ ) zones of inhibition compared to vehicles and untreated controls. Inhibition zone areas against MRSA USA300 and AB19606 ranged between 30-43 cm<sup>2</sup>.

**Discussion:** Our results demonstrated that MOX can be loaded into the topical exosome-loaded gels and has significant antimicrobial activity against two common wound pathogens. These results warrant future preclinical and clinical studies to substantiate their use in preventing or treating wound infections.

## Introduction

Delayed wound healing is frequently associated with wound infections.<sup>1</sup> Resolving these infections is made difficult by drug resistant pathogens such as the ESCAPE pathogens *S. aureus* and *A. baumannii*.<sup>2</sup> Methicillin Resistant *S. aureus* produces the greatest burden among notable MDR organisms on the healthcare system, with estimated cost in the United States of \$1.2 billion annually. Carbapenem resistant *Acinetobacter* infections produce the greatest cost per community acquired infection.<sup>3</sup> This study investigated the efficacy of a novel encapsulated antibiotic against MRSA and *A. baumannii* clinical isolates through a tube dilution test and a modified Kirby-Bauer assay.

**References**

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### 1. Microorganisms:

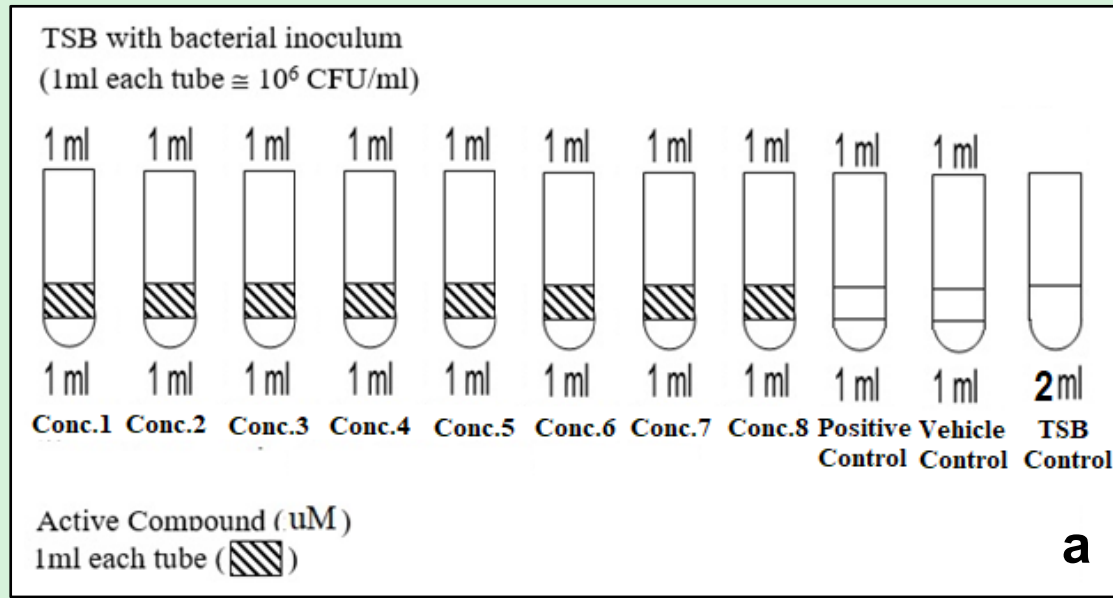
- Fresh cultures of Methicillin Resistant *Staphylococcus aureus* (MRSA) USA300 and *Acinetobacter baumannii* ATCC 19606 (AB19606) were used in this study.

### 2. In vitro Studies:

- This study was conducted with several phases or tasks. First, minimum inhibitory concentrations of Moxifloxacin against MRSA and AB19606 were determined.
- The second task investigated the zone of inhibition produced after encapsulation of MOX into hybrid exosomes at varying concentrations compared to unencapsulated
- The third task investigated change in efficacy after incorporation of the MOX-loaded hybrid vesicles into gels for potential topical application.
- The fourth task was conducted to evaluate efficacy after varying storage temperatures and times.

### 3. Minimum Inhibitory Concentration (MIC)

- Overnight growth of bacterial cultures was suspended in sterile tryptic soy broth (TSB).
- 1 mL of challenge inoculum suspension was combined with 1 mL of treatment per tube for a concentration of approximately 10<sup>6</sup> CFU/mL. 3 replicates were performed per treatment (a).
- Tubes were incubated for 24 hours at 37°C.
- Degree of turbidity was assessed, and bacterial concentration was quantified by plating serial dilutions of each replicate onto tryptic soy agar (TSA) plates.
- The plates were incubated overnight at 37°C and CFUs were tabulated to determine the MIC.

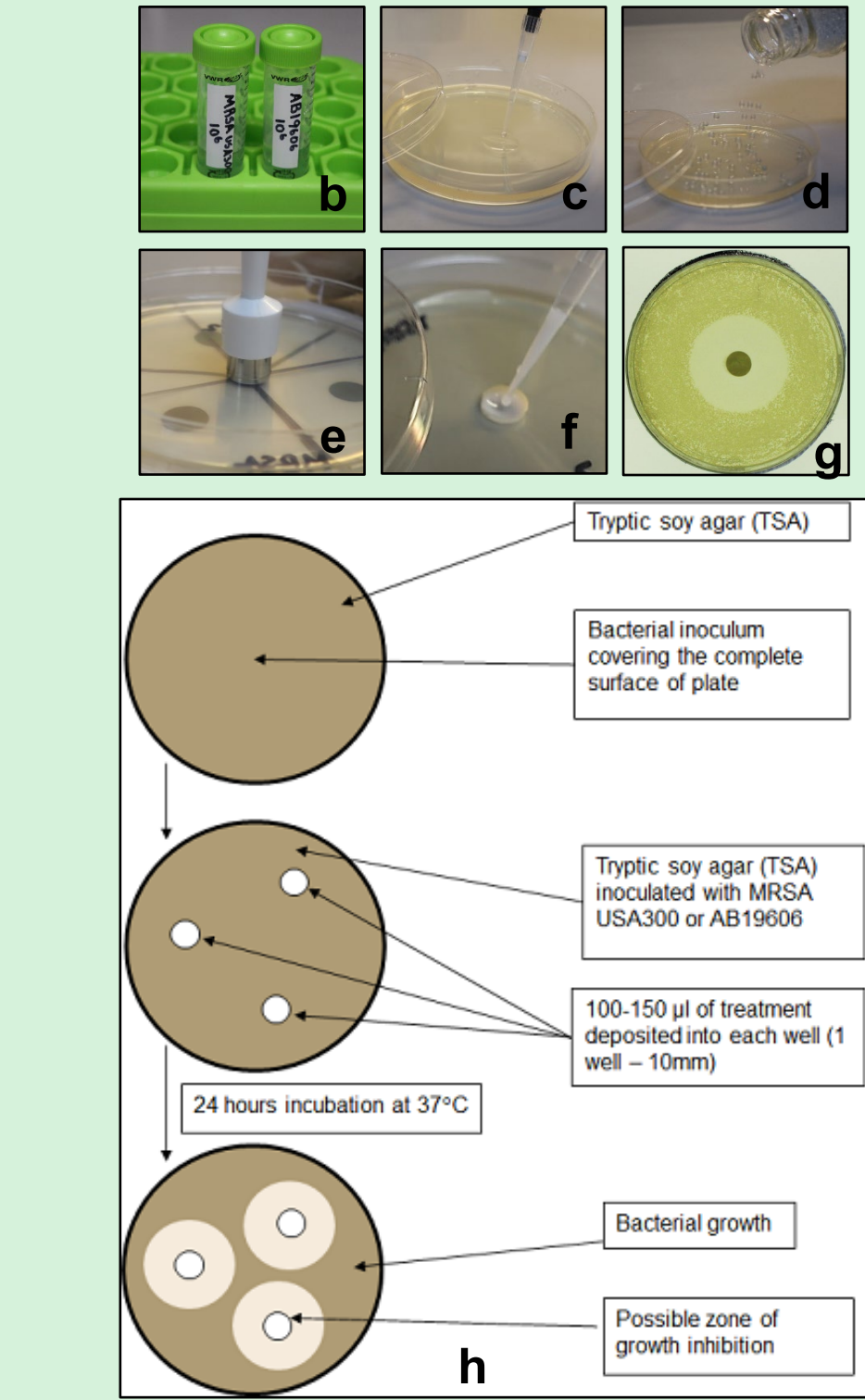


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## Materials and Methods

### 4. Zone of Inhibition Assays

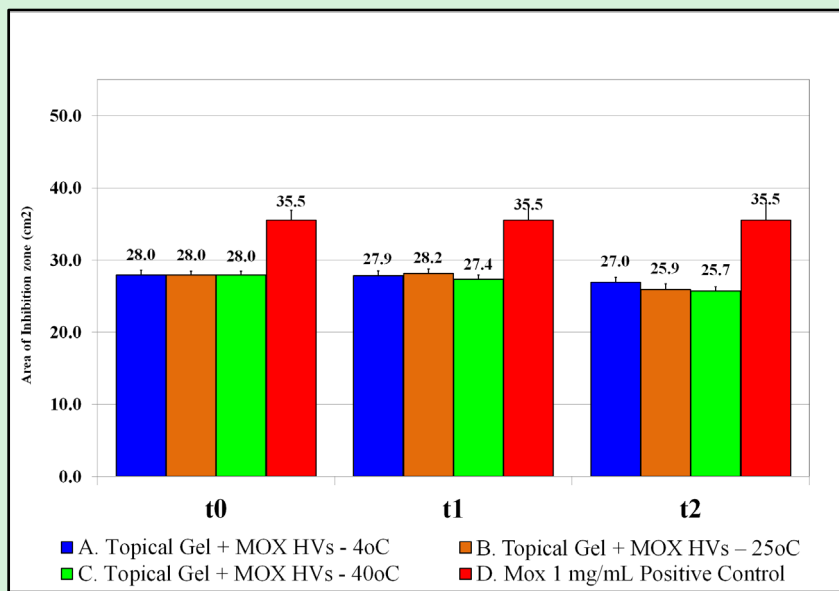
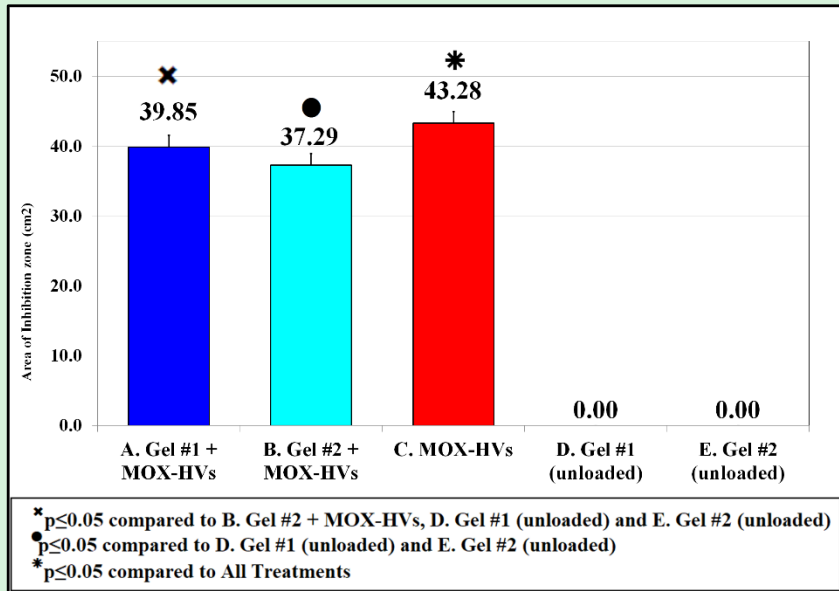
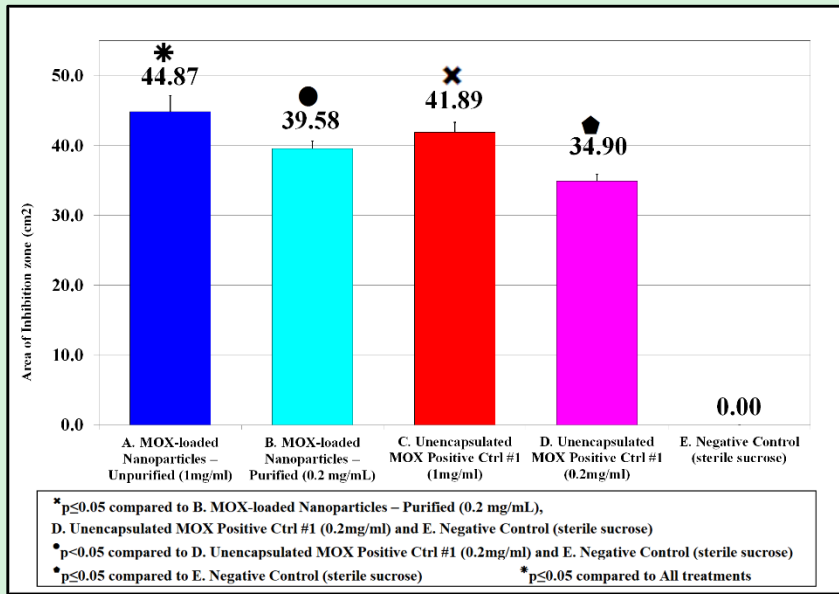
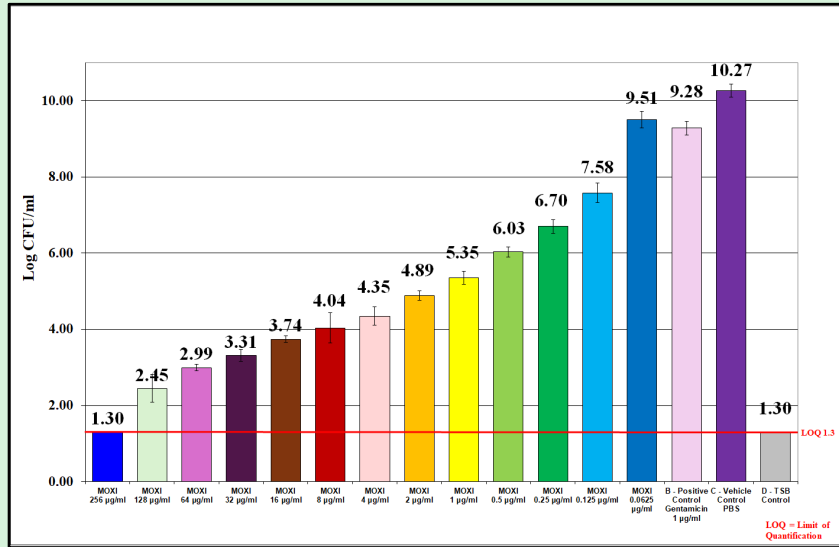
- A challenge inoculum with a concentration of 10<sup>6</sup> CFU/mL was prepared by scraping overnight growth of bacterial cultures into TSB (b).
- 100 µL of this inoculum was deposited onto TSA plates and spread evenly with sterile glass beads (c)(d).
- Three punch biopsies (10 mm) were made per plate with the agar within removed to produce wells (e).
- 100-150 µL of each treatment was deposited into these wells (f).
- Treatments was allowed 2-3 hours for diffusion, the plates then were incubated for 24 hours at 37°C (see diagram h).
- After incubation, zones of bacterial growth inhibition were measured using ImageJ 1.41o software, below example (g).



### 6. Statistical analysis

- Statistical analysis was conducted using one way ANOVA IBM SPSS v29 was performed in Task#2 and Task#3. For Tasks 1 and 4 statistical analysis was not performed.

### MRSA USA300



## Conclusions

- The loading of MOX into exosomes enhanced the antimicrobial efficacy of the antibiotic. The MOX-loaded nanoparticles produced greater zones of inhibition than unencapsulated MOX at identical concentrations and remain effective in a topical gel vehicle or at various storage timepoints and temperatures.
- MOX-loaded hybrid vesicles are a promising treatment for wound infections by both Gram-positive and Gram-negative pathogens.

## Results

### Task #1 MIC Results

- The MIC for unencapsulated MOX against MRSA was 256 µg/mL. Against AB19606, the MIC was 32 µg/mL.
- The MICs observed reduced bacterial growth below the limit of quantification.
- Positive Controls, Gentamicin (1 µg/mL) against MRSA USA 300 or Silver Sulfadiazine (30 µg/mL) against AB19606, showed 9.28 ± 0.18 and 9.87 ± 0.35 Log CFU/mL, respectively.
- Vehicle control against MRSA USA300 and AB19606 exhibited the highest bacterial 10.27 ± 0.17 and 10.25 ± 0.20 Log CFU/mL, respectively.

### Task #2 Encapsulation Results

- Against MRSA, the encapsulated or MOX-loaded nanoparticles produced inhibition zones at both concentrations tested that were significantly ( $p \leq 0.05$ ) greater in diameter compared to zones of unencapsulated MOX.
- The MOX-loaded Nanoparticles at 0.2 mg/mL had significantly ( $p \leq 0.05$ ) larger zones of inhibition (ZOI) against AB19606 than the unencapsulated MOX at 0.2 mg/mL. The greatest inhibition was observed against MRSA with the 1 mg/mL MOX-loaded nanoparticles producing a ZOI of 44.87 cm<sup>2</sup>.

### Task #3 Gel Vehicle Results

- The MOX-loaded hybrid vesicles (MOX-HVs) in Gel #1 against MRSA produced ZOIs that were significantly ( $p \leq 0.05$ ) larger than those for Gel #2 with Mox-HVs and the Vehicle Control Gels #1 and #2.
- The MOX-HVs in Gel #1 had ZOIs against AB19606 that were larger than those for MOX-HVs in Gel #2 though this difference was not significant.
- The largest ZOIs against both pathogens was observed for treatment group C, with ZOIs of 43.28 cm<sup>2</sup> against MRSA and 36.41 cm<sup>2</sup> against AB19606. These were statistically significant ( $p \leq 0.05$ ) compared to all other treatment groups for both pathogens.

### Task #4 Storage Viability Results

- The ZOIs produced by the MOX HVs in Topical Gel at time point 2 (t2 or two weeks) and stored at 4°C were similar to those produced at t0 also stored at 4°C against both pathogens.
- The ZOI for the storage at 40°C was smaller (25.7 cm<sup>2</sup>) at t2 against MRSA USA300 compared to storage at 4°C (28.0 cm<sup>2</sup>) but against AB19606, the ZOI after 40°C at t2, 25.7 cm<sup>2</sup>, is more similar to t0 (25.8 cm<sup>2</sup>).
- Positive Control MOX 1 mg/mL showed against both organisms similar ZOI (35.5 and 34.2 cm<sup>2</sup>, respectively).

### AB19606

