Young-Joon Jun Department of Plastic Surgery, The Catholic University of Korea

Introduction

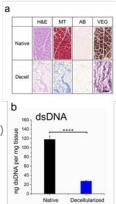
Volumetric muscle loss (VML) is an irrecoverable injury generally more than 20% muscle loss. The decellularized extracellular matrix (dECM) approach has been extensively investigated. In this study, we developed pre-vascularized muscle tissue constructs for VML treatment using muscle dECM (mdECM) bioink through 3D cell printing. The regenerative capability of muscle tissue constructs was evaluated through implantation in a rat model of the VML defect.

Materials and Methods

Human muscle cells and human umbilical vein endothelial cells (HUVECs) were encapsulated in mdECM and vascular-derived dECM (vdECM) bioink. Pre-vascularized skeletal muscle constructs were fabricated in the gelatin granule reservoir. To create the models of VML injuries, 10-week-old Sprague Dawley rats were anesthetized, and 40% of the tibialis anterior muscle was removed from each rat. The 3D cell printed muscle constructs were implanted and sutured at both ends. Masson's Trichrome staining, immunofluorescence staining, and functional assessment were conducted at four weeks post-impanation.

dECM bioink & cell preparation

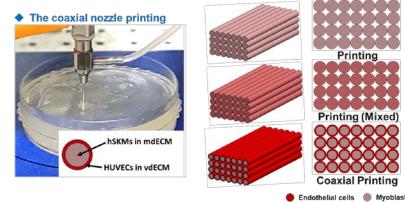
- Skeletal muscle dECM (mdECM)
 porcine tibialis anterior (TA) muscle
- Vascular dECM (vdECM)
- porcine descending aortas
- Human skeletal muscle cells (hSKMs)
- Human umbilical vein endothelial cells (HUVECs)



27.57 ± 1.31 ng of dsDNA

per mg of tissue

Fabrication of vascularized muscle construct



- Vascularized skeletal muscle construct
 - Two types of cells and bioinks were considered for fabrication of vascularized muscle.
 - > Coaxial nozzle printing was applied



mplant the 3D cell printed muscle construct

Excise approximately 40% (mass) of the rat TA muscle to create VML.

TA muscle wt = 0.0017 x body wt = 0.0715

Remove EDL and EHL muscles to rule out compensatory hypertrophy.

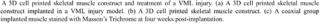
3D Cell Printed Muscle Tissue for Functional Muscle Recovery

Young-Joon Jun Department of Plastic Surgery, The Catholic University of Korea

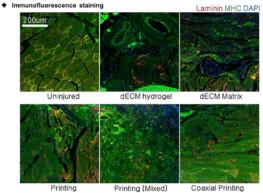
Results

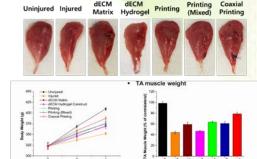
In vitro assessments showed that the pre-vascularized muscle tissue construct that was formed by a coaxial nozzle (coaxial group) exhibited enhanced contractile force and myotube and endothelial network formation. In vivo results revealed that the muscles treated with the coaxial group greatly improved de novo muscle fiber regeneration, vascularization, and innervation. Most importantly, the pre-vascularized muscle tissue construct achieved 85% functional recovery in VML injuries.

a b c a b c A 3D cell printed skeletal muscle construct and treatment of a VML injury. (a) A 3D cell printed skeletal muscle



Treatment of volumetric muscle loss injury





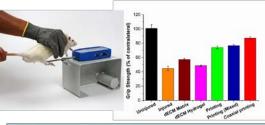
Conclusion

In this study, we developed a pre-vascularized muscle construct using 3D cell printing and a coaxial nozzle. We expect that these results will provide a blueprint for the future development of engineered human-scaled muscle tissues

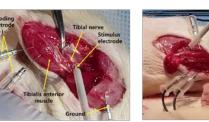
for VML injury treatment.

Treatment of volumetric muscle loss injury

Results (Grip strength)



Functional assessment by grip strength measurement
 The injured muscles showed ~55% reduction in grip strength with unijured muscles.
 The coaxial printing group reached ~87% of functional recovery



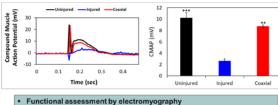
Treatment of volumetric muscle loss injury

Results (Electromyography)

Functional assessment by electromyography
 TA muscles were exposed and electrodes were inserted.
 Stimulation was applied to the Tibial nerve.

Treatment of volumetric muscle loss injury

Results (Electromyography)



The injured muscles showed -74% reduction in CMAP compared with uninjured muscles.
 The coaxial group showed a 3-fold improvement in CMAP

 Ine injured muscles showed ~ compared with uninjured musc
 The coaxial group showed a 3compared to the injured group.