

Engineered Nanobubbles for Inducing Unprogrammed Necrosis in Cancer Immunotherapy

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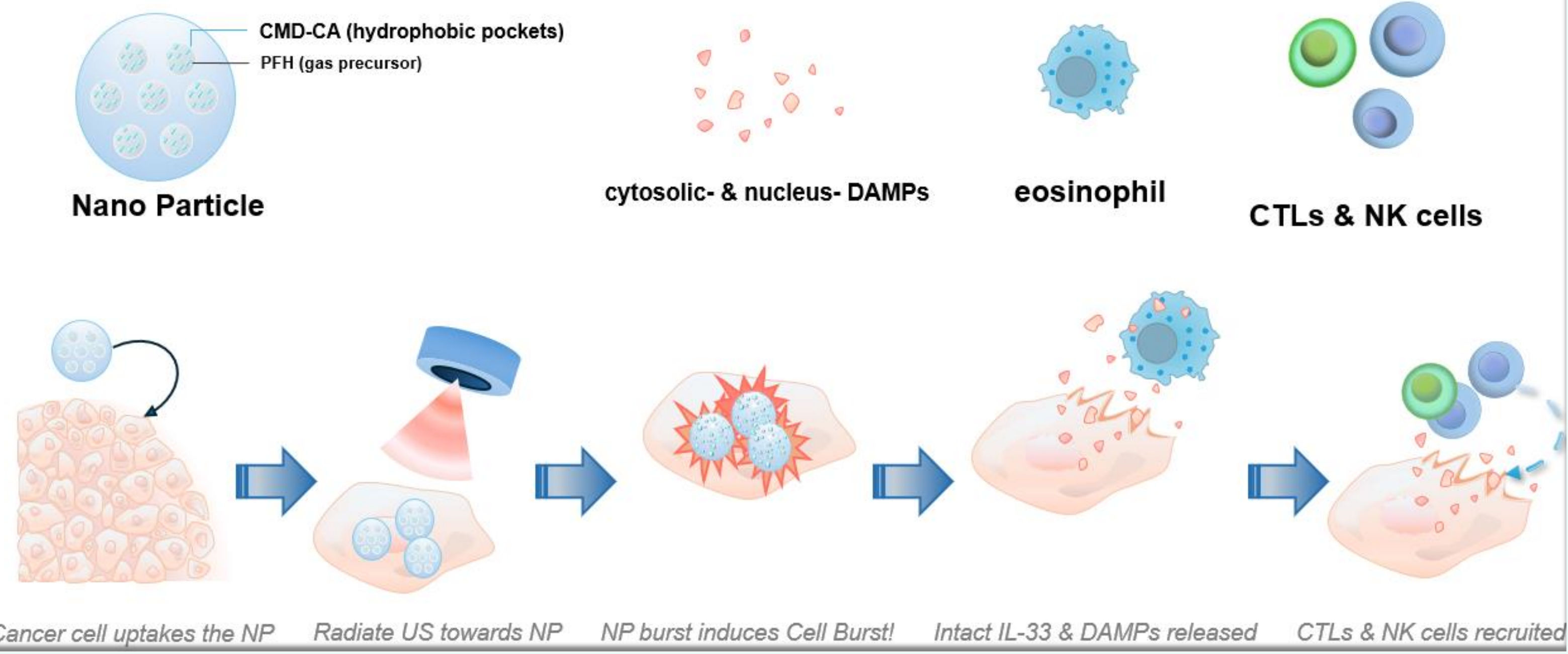
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KEY WORDS: IL-33, eosinophils, unprogrammed cell death, CTLs and NK cells recruitment

Abstract

Unprogrammed cell deaths play a key role in the modulation of inflammatory responses by releasing various immunostimulatory molecules. In particular, accidental necrosis can activate eosinophils by releasing interleukin-33 (IL-33) and thereby provide an exceptional opportunity for the recruitment of natural killer cells and cytotoxic T-cells in the antitumor immune response. In this research, we designed nanobubbles that are cavitated by ultrasound to induce cell death, which also dramatically leaks IL-33 and damage-associated molecular patterns (DAMPs).

Scheme



Conclusion

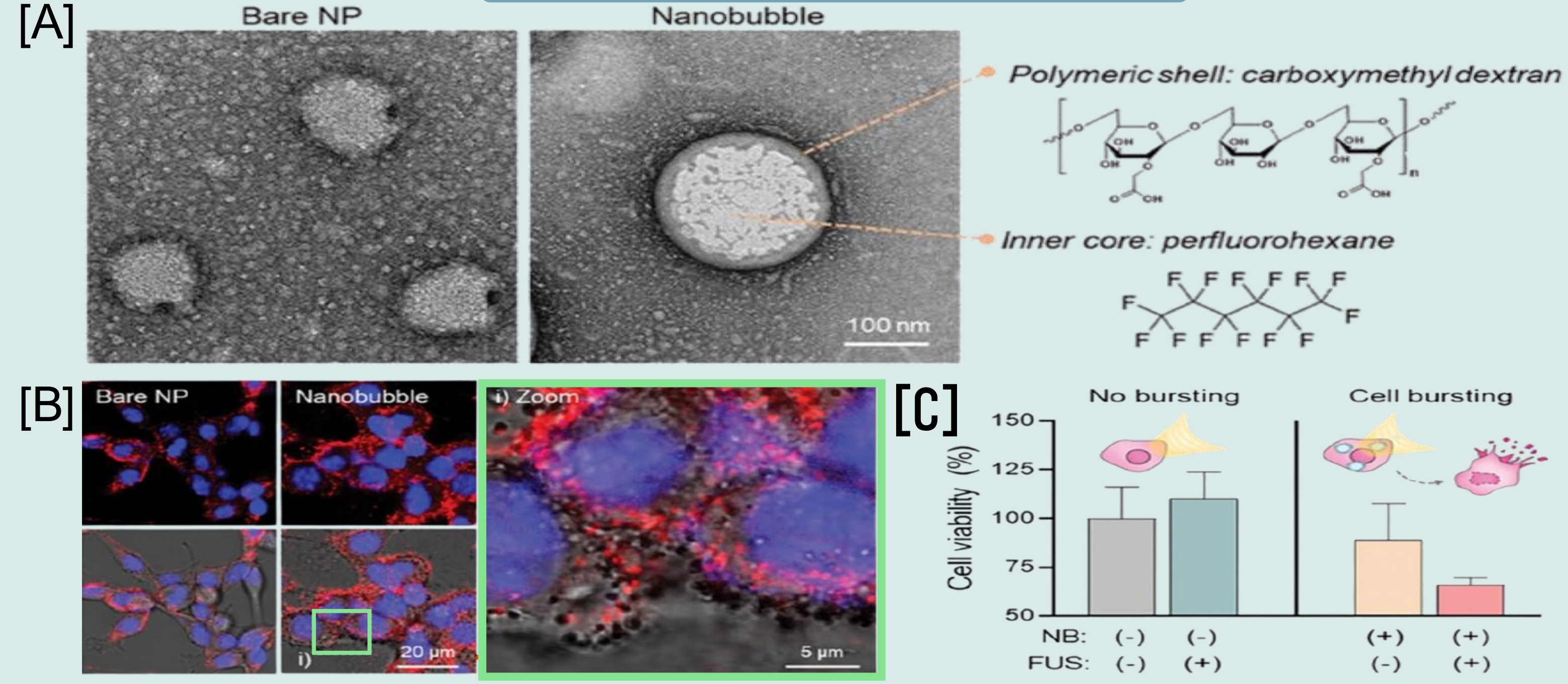
NanoBurst triggers complete **IL-33 & DAMP** release via ultrasound-induced cell rupture
Released DAMPs → **Eosinophil activation**
→ **Robust CTL/NK cell recruitment** at tumor site
with DPP4i combination shows [CD69⁺ (2.66×) ↑, CD8⁺ (1.46×) ↑] in lung
(Stronger than NanoBurst + aPD-1)
• **NanoBurst-driven CTL/NK recruitment fuels potent antitumor immunity!**



References!

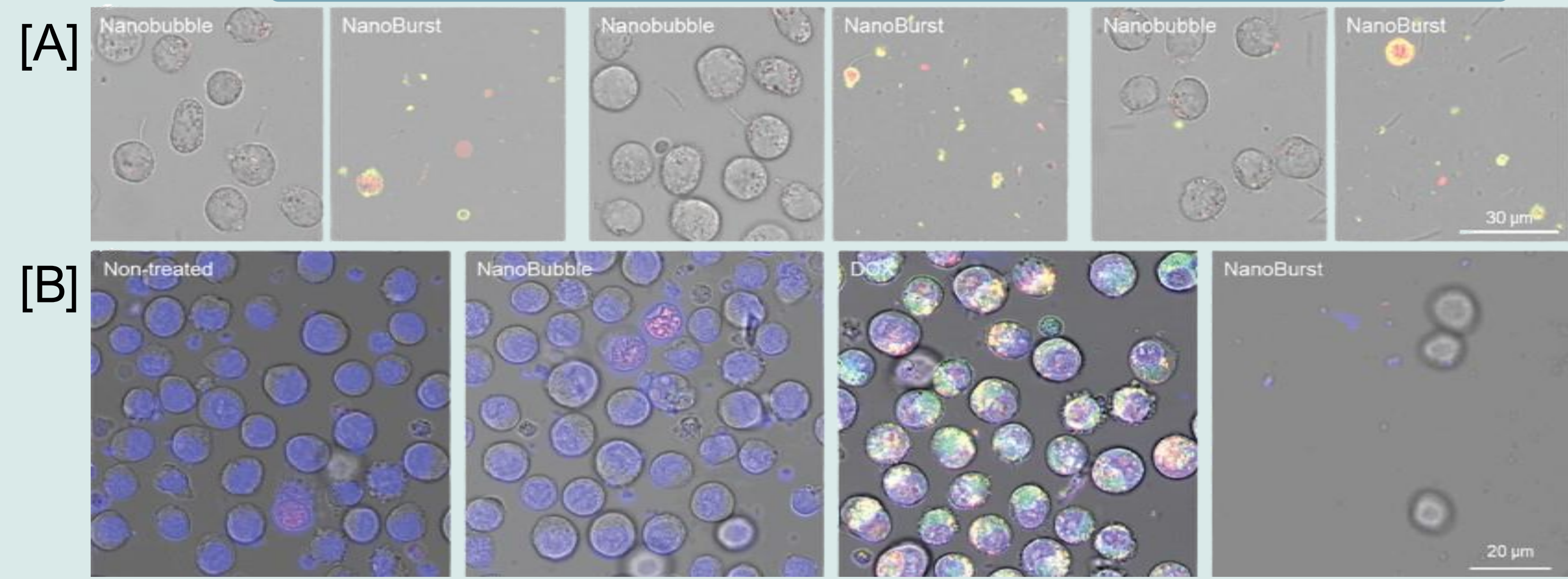
Results

Fig.1 Characterization of NPs



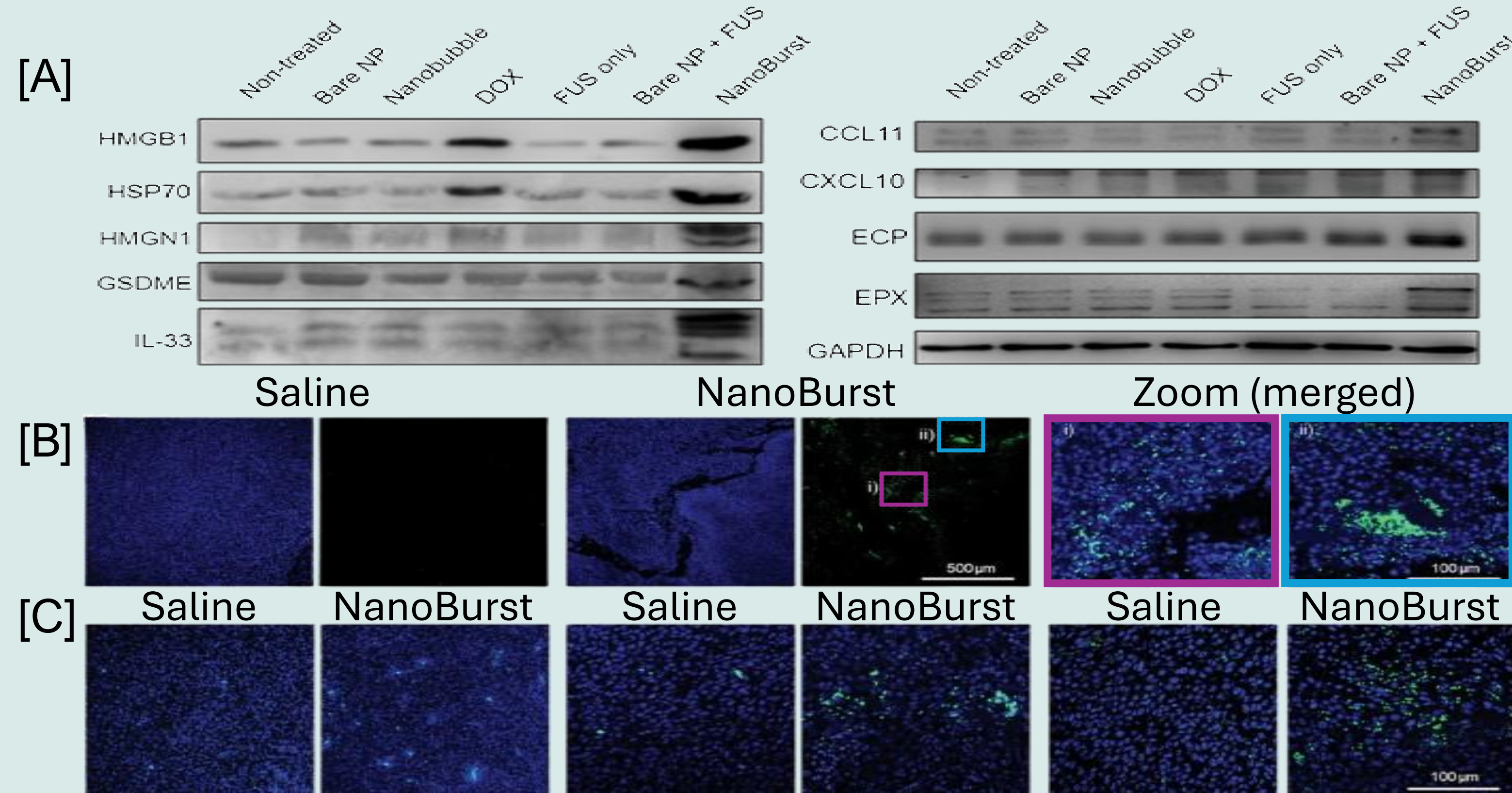
[A] TEM images and the chemical structures of nanobubble components. [B] Confocal microscopic images of CT26 cells treated with (left) bare NPs or (right) nanobubbles (100µg/mL) for 6h. (DAPI & Cy5.5) [C] Cell viability of CT26 cells treated with ultrasound, (left) nanobubbles, or (right) NanoBurst.

Fig.2 Different cell death type from Doxorubicin



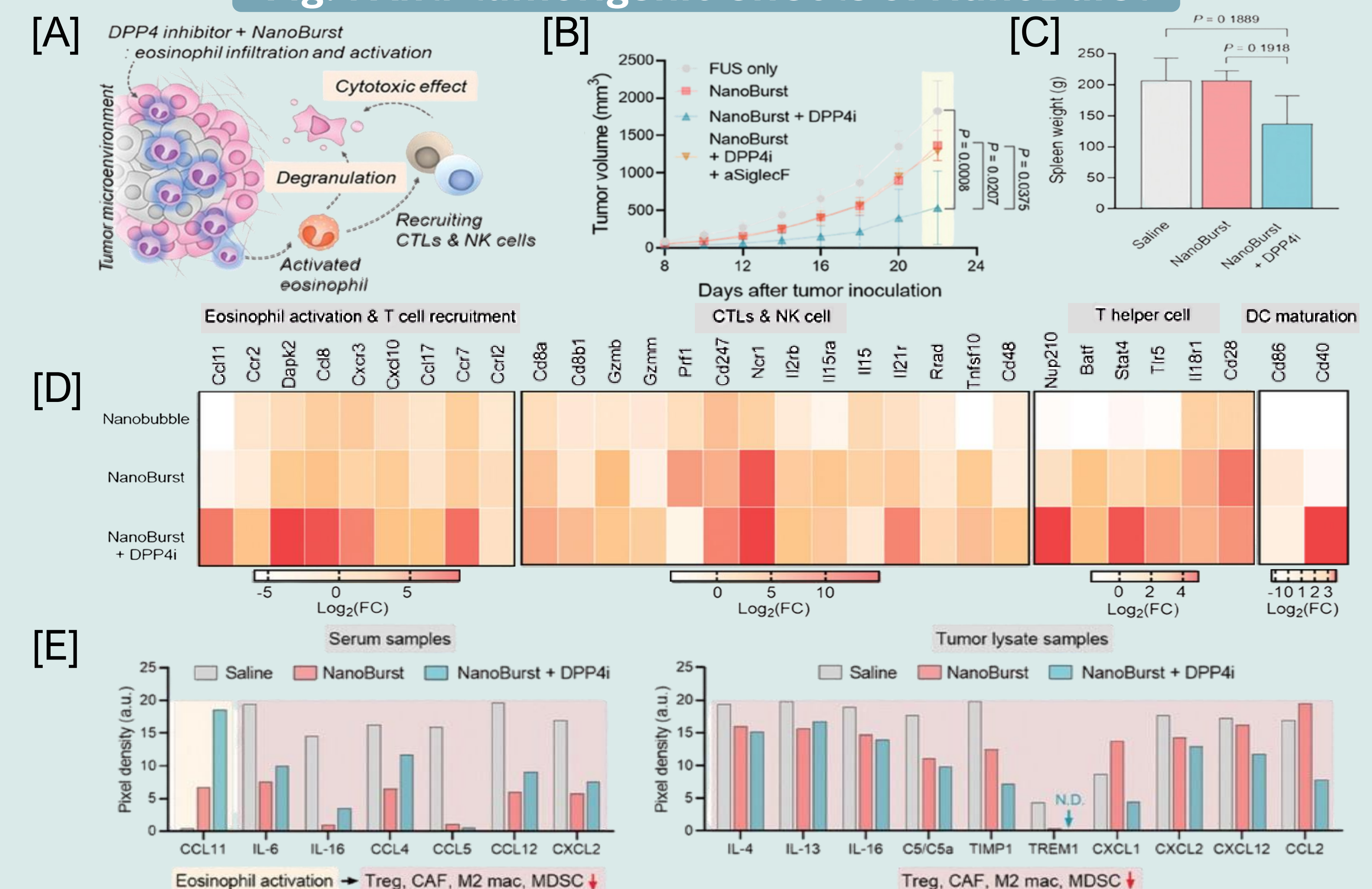
[A] Annexin V/PI imaging of MLKL knockdown CT26 cells treated with (left) nanobubbles or (right) NanoBurst. [B] Caspase-3 imaging (DAPI, DOX, Cas3) of CT26 cells treated with [non-treated | nanobubbles | DOX | NanoBurst].

Fig.3 Intact DAMP release via NanoBurst



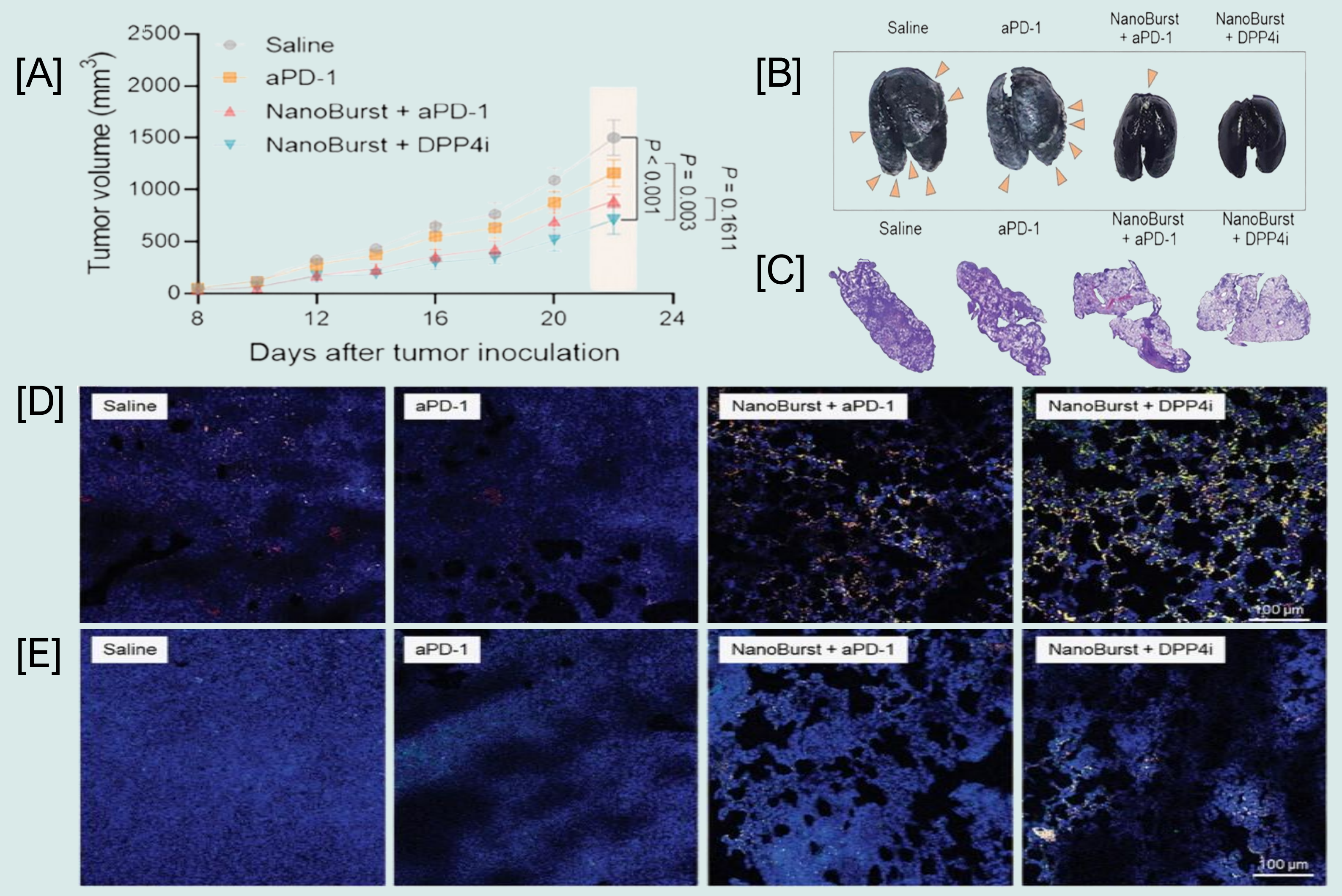
[A] Western blot of the conditioned medium treated with [non-treated | bare NPs | nanobubbles | DOX | US | bare NPs + US | NanoBurst] (left) from CT26 cells, (right) from eosinophils after incubation with the CT26 cells. [B] Immunohistochemistry of activated eosinophils in tumor tissues. (DAPI & SiglecF) [C] Immunohistochemistry of DAPI & CTLs, NK cells, and matured DCs in tumor tissues.

Fig.4 Anti-tumorigenic effects of NanoBurst



[A] Schematics for anti-tumorigenic effects. [B] Changes in tumor volume for each treatment group (n=4). [C] Weight of the spleens for each treatment group (Saline, NanoBurst, NanoBurst+DPP4i). [D] Heat map of mRNA levels in tumor tissues for each treatment group (Nanobubble / NanoBurst / NanoBurst+DPP4i; downwards). [E] Cytokine and chemokine levels for each treatment group (Saline, NanoBurst, NanoBurst+DPP4i).

Fig.5 Synergic effects of NanoBurst + DPP4i



[B]~[E]: [saline | aPD-1 | NanoBurst + aPD-1 | NanoBurst + DPP4i]
[A] The average tumor volume. [B] Representative photographs of India-ink-stained lung tissues. [C] Hematoxylin and eosin staining images of the lung tissues. [D] Immunohistochemistry of activated eosinophils. (DAPI, SiglecF, CD69) [E] CTLs in the lung tissues. (DAPI, CD3, CD8)