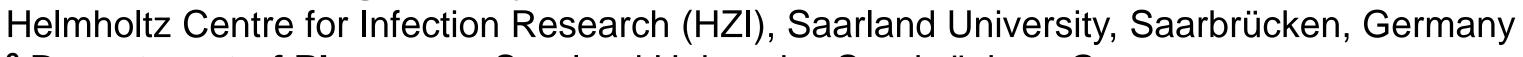
Acid-cleavable Tobramycin cross-linked nanogels for antibiotic delivery to P. aeruginosa biofilms



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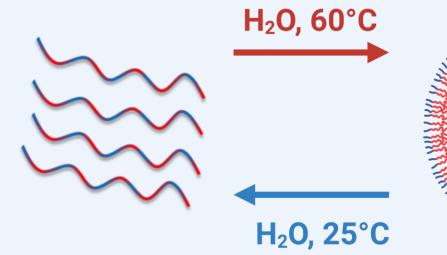
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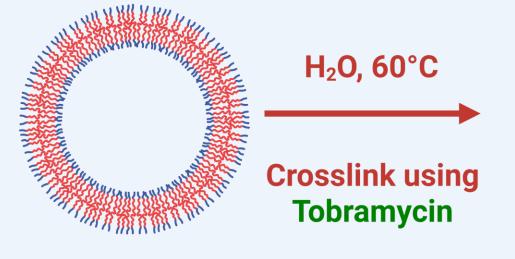
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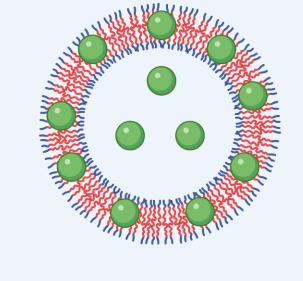
Synopsis

Biofilm (BF) formation significantly reduces the susceptibility of *Pseudomonas* aeruginosa (PA) to Tobramycin (Tob) and other aminoglycosides (AGs). This is mainly due to the electrostatic interactions between cationic AGs and anionic species within the BF (1), especially at acidic pH. We hypothesized that nanogels (NGs) (2) can shield the cationic charge of AGs and therefore enhance their BF penetration. To produce such NGs, a new diblock copolymer was synthesized. The first block of the copolymer comprises linear PEG. In the second block, a thermoresponsive poly(oligoethylene glycol) methacrylate (POEGMA) and an aldehyde functional monomer are randomly distributed.

To create NGs, the polymer was cross-linked using Tob in water above its lower critical solution temperature (LCST) via imine bonds. The NGs were found to be biocompatible, rapidly released Tob at pH ≤ 6.0, as reported for PA BF (3), and killed planktonic PA. Targeted Tob release in infected or inflamed areas ensures bacterial killing primarily at those sites. This approach may reduce collateral damage to the microbiome and potentially slow the development of AMR. However, in biofilms the killing is insufficient, likely both due to PA having decreased susceptibility to Tob in acidic conditions, as well as the general susceptibility decrease observed in biofilms.

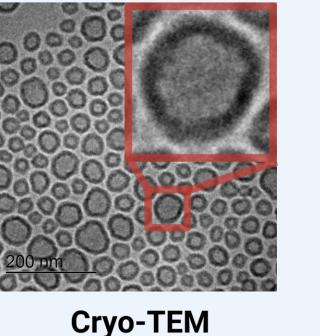


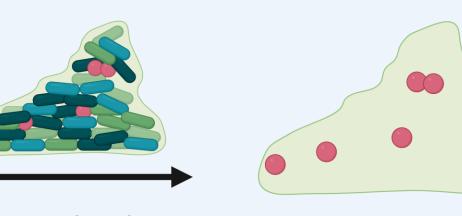




Cool down

to RT





Penetration into PA BF

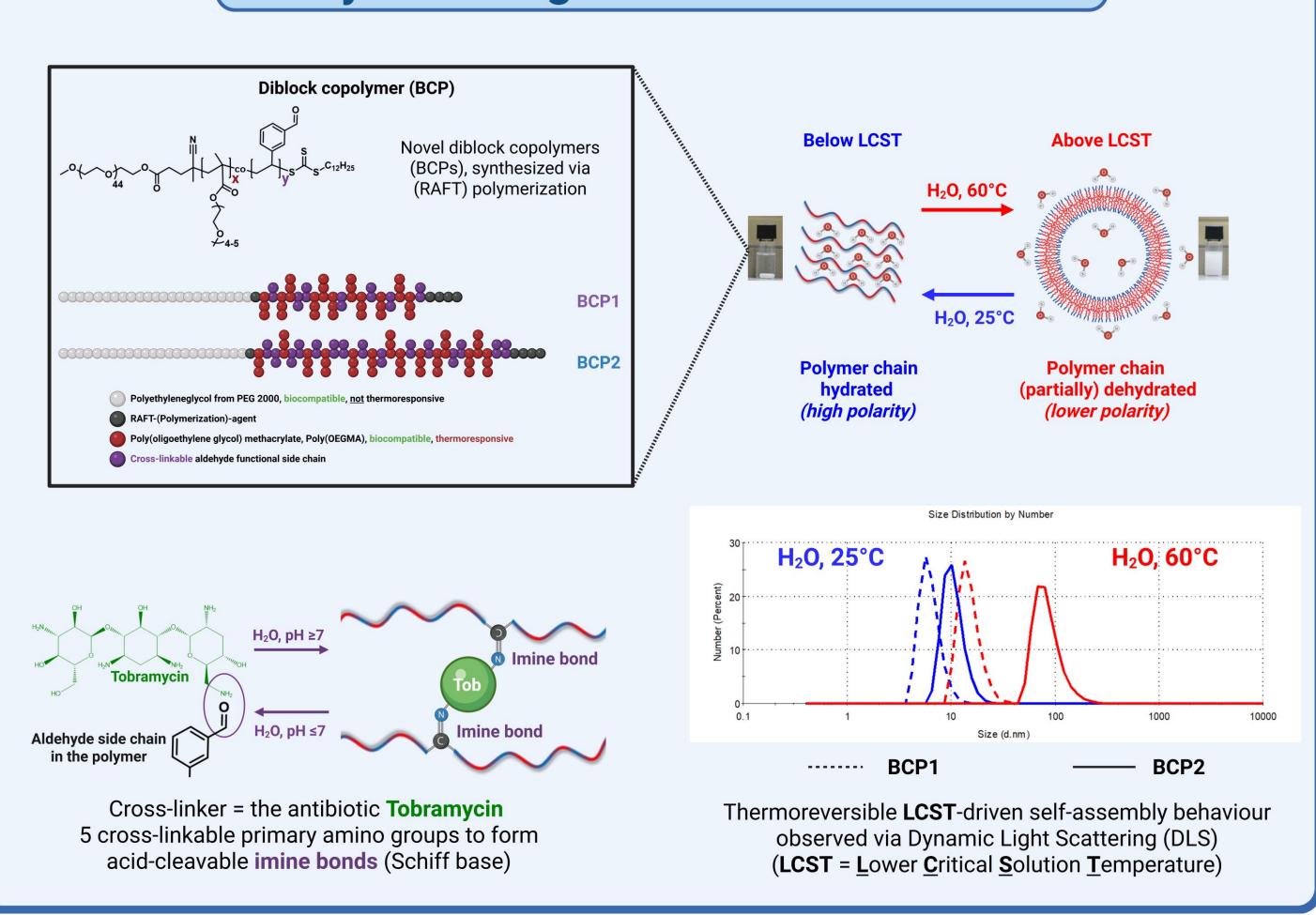
Acidic pH-mediated Tob release & bactericidal effect within the BF

Solvated tailor-made block-copolymers

Self-assembled nanogel (thermoreversible)

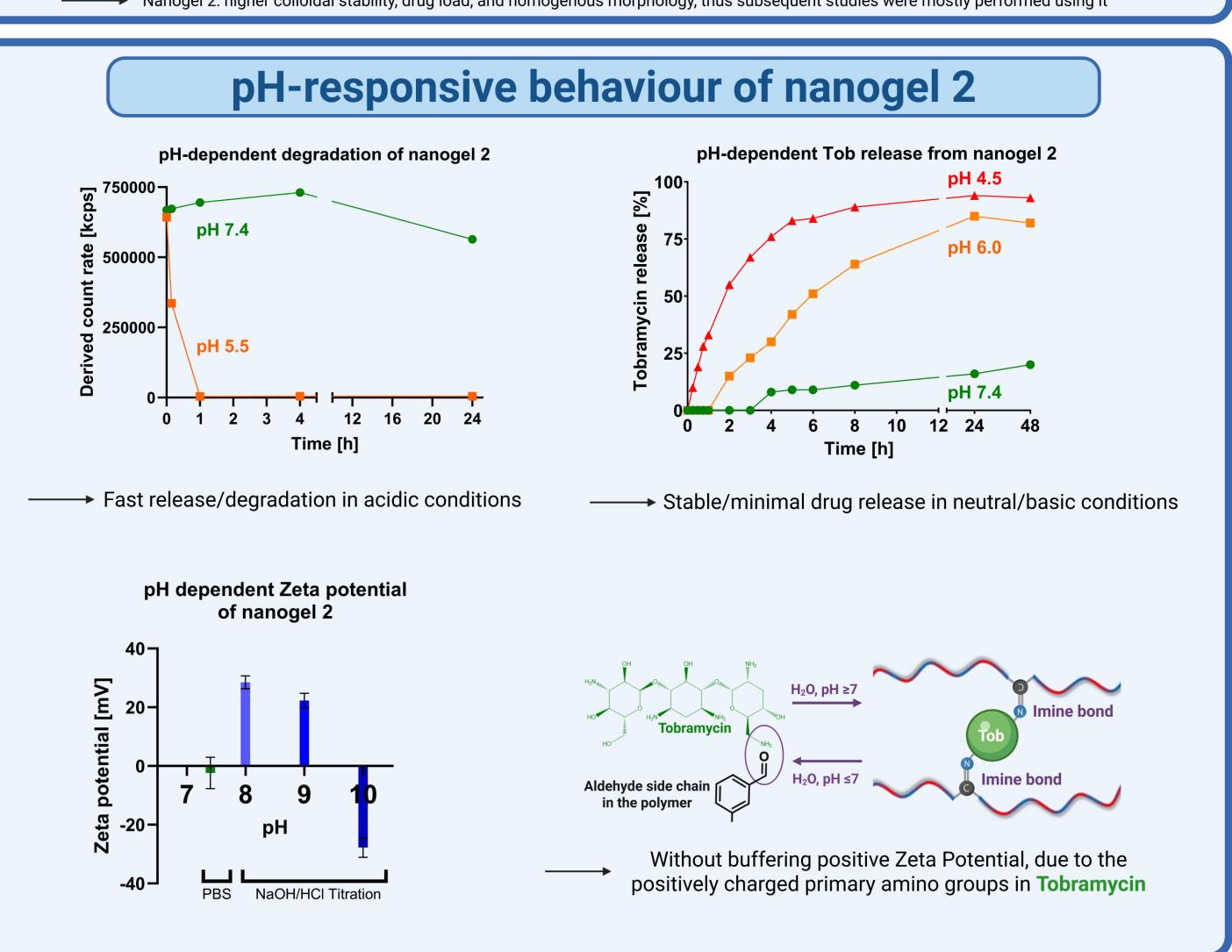
Tob cross-linked nanogel (Tob-polymer conjugate)

Polymer design and characterization



Nanogel preparation and characterization **Loading rate / Encapsulation Efficiency Nanogel preparation** Cross-link 60 °C, 300rpm 2 - 24 h **Cryo-TEM** Nanogel 2 resulting from BCP2 shows homogenous vesicles Nanogel 1 resulting from BCP1 shows micelles Nanogel 1 from BCP1 Nanogel 2 from BCP2 Intensity based Size Distribution of nanogels via Dynamic Light Scattering (DLS) Encapsul. Efficiency Loading rate 11% Conc. of loaded Tob 1209 1141 **Z-Average** 23.4 ± 0.3nm 85.7 ± 2.8 nm 0.155 ± 0.009 0.085 ± 0.016 Zeta potential 0.19 ± 0.18 mV -0.52 ± 1.29 mV Nanogel 2: higher colloidal stability, drug load, and homogenous morphology, thus subsequent studies were mostly performed using it

Cytocompatibility and bactericidal effect A549 cells Optical image analysis of alive and dead cell count using TECAN Image Analyzer Software MIC/MBC/MBEC of Tobramycin and Nanogel 2 for Pseudomonas aeruginosa (PAO1) MBC in LB Broth [µg/mL] MIC90 in LB Broth [µg/mL] Standard Conditions pH 5 pH 5 Free Tobramycin Tob nanogel 2 N=2 MIC = Minimum Inhibitory Concentration, MBC = Minimal bactericidal concentration n=4 PAO1 was incubated in for 72h to form biofilms → treated for 48h with Tob or NG A549 cells treated for 24h with highest nanogel 2 concentration Release kinetic issue in bacterial killing? → pre-release experiment: Tob and nanogel 2 (256µg Tob/mL; 1,5mg BCP2/mL) pre-incubated at pH 6 37°C for 72h before used in treatment stained with LIVE/DEAD™ Kit (Calcein-AM / ethidium homodimer-1) → same negligeable decrease in CFU like in the pH 6 assay Biofilm (pH 7-8) treated Biofilm (pH 6) treated A549 Viability after 24h treatment A549 Viability after 24h treatment with Tobramycin vs nanogel 2 with Tobramycin vs nanogel 2 Tob XX Tobramycin conc. of Polymer concentration [mg/mL] BCP 1



Conclusion

- Using a novel, thermoresponsive diblock copolymer with aldehyde functionality, we formulated acid-cleavable Tob-crosslinked nanogels
- Drug loading contents of 1300 µg/mL while a PEG-shell effectively shields the positive charge of Tobramycin
- Promising new pH-controlled release system for prolonged release of Tob in antibacterial therapies
- Maintains the antibiotics intrinsic activity even after nanoencapsulation
- Adhesion of the nanogels to biofilms (infected lung or topical/burn wounds) -> longer residence and selective release only in infected environments → combatting antimicrobial resistance (AMR)

Outlook

- Co-delivery by loading/conjugation of another antibiotic or adjuvant with antimicrobial properties
- Use outside of infection research with other drugs possessing multiple primary amino groups, such as peptides

References

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- (3) Hollmann, B., Perkins, M., Chauhan, V. M., Aylott, J. W., & Hardie, K. R. (2021). Fluorescent nanosensors reveal dynamic pH gradients during biofilm formation. NPJ Biofilms Microbiomes, 7(1), 50. https://doi.org/10.1038/s41522-021-00221-8