HA-PEG Layered F-PEI Core-Shell Micelles for PDGFRβ Targeted Delivery of TXNDC5 Silencing for Pulmonary Fibrosis

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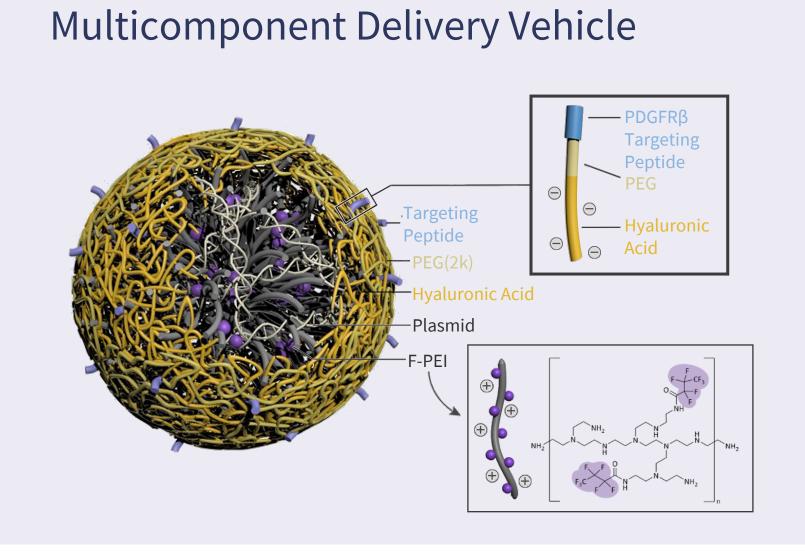
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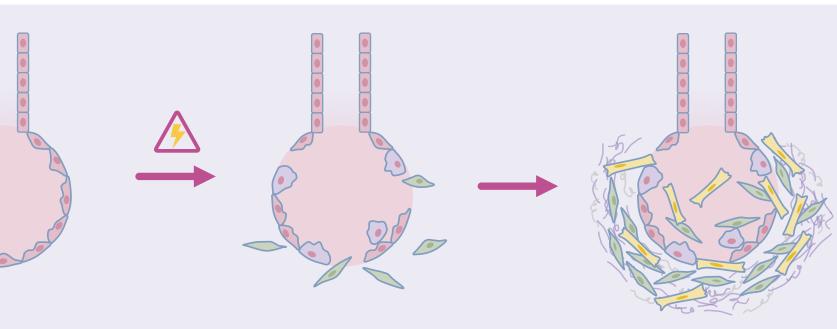
Highlights

- Multifunctional core–shell design enhances stability, circulation, and targeted shRNA delivery
- F-PEI self-assembles with shRNA to form compact, fully condensed cores
- PDGFRβ-targeted delivery improves uptake in fibrotic lung cells
- HA-PEG shelling retains endosomal escape and ensures high cell viability
- Therapeutic efficacy in bleomycin model with reduced fibrosis and marker expression



Disease Background

Idiopathic pulmonary fibrosis (IPF) is a fatal lung disease marked by progressive, irreversible scarring. IPF involves chronic inflammation, fibroblast activation, and excess ECM deposition. Current therapies slow progression but do not reverse disease, highlighting the need for targeted treatments.

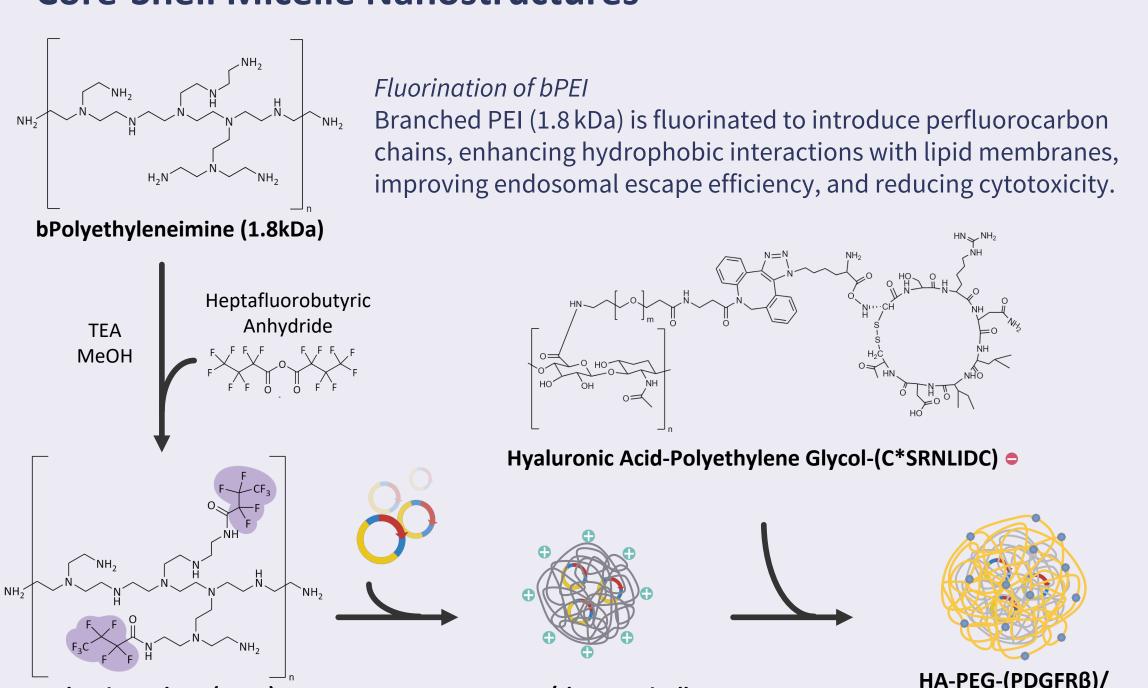


Healthy Alveola
Intact epithelium and basement
membrane support gas
exchange and lung elasticity.

Injury & Inflammation
Epithelial damage attracts
immune cells that release
cytokines driving fibrosis

Aberrant Wound Repair
Injury activates fibroblasts,
leading to ECM buildup and
irreversible lung scarring

Synthesis and Assembly of shRNA/F-PEI-HA-PEG-(PDGFRβ) Core-Shell Micelle Nanostructures



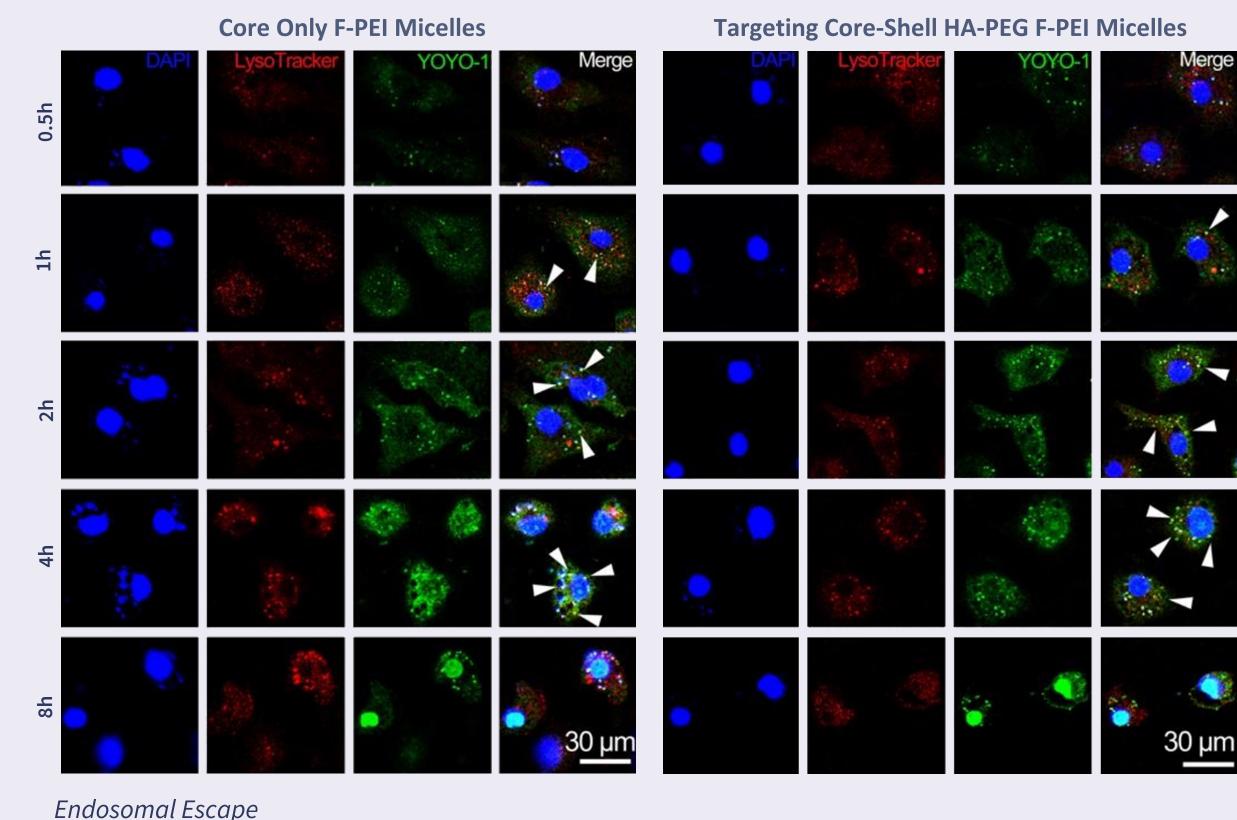
shRNA Complexation
Negatively charged shRNA rapidly
self-assembles with cationic F-PEI via
electrostatic interactions to form compact
core micelles.

Fluorinated PEI (F-PEI)

Targeted Core-Shell Nanostructure
A HA-PEG shell decorated with
PDGFRβ-targeting peptide is grafted onto the
F-PEI/shRNA core, imparting colloidal
stability and receptor-mediated targeting

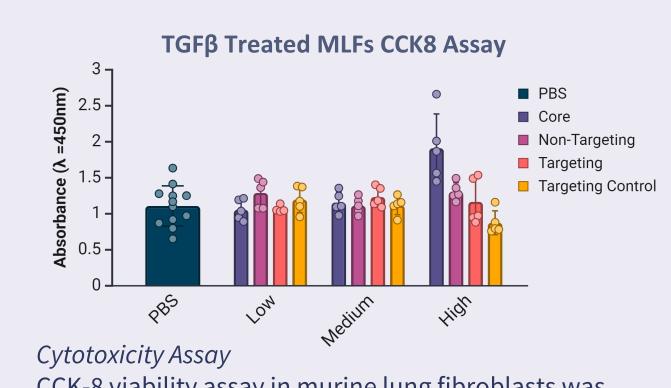
F-PEI/shRNA Micelle

Core-Shell Structure Retains Effective Endosomal Escape

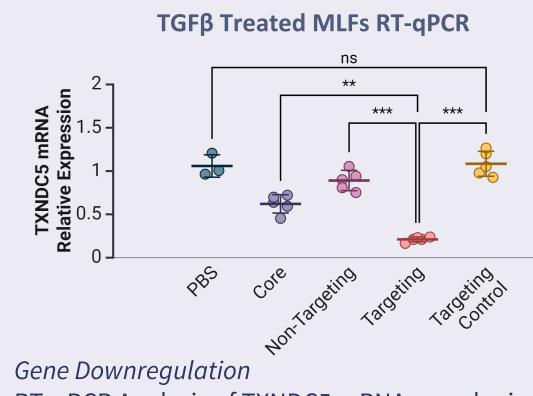


Lysosomal trafficking assessed by colocalization of YOYO-1-labeled shRNA (green) with LysoTracker (red) indicates both core and core-shell particles achieve endosomal escape and nuclear accumulation. The HA-PEG shell slightly slows escape kinetics but does not abrogate delivery efficacy.

Cell Viability & TXNDC5 Knockdown



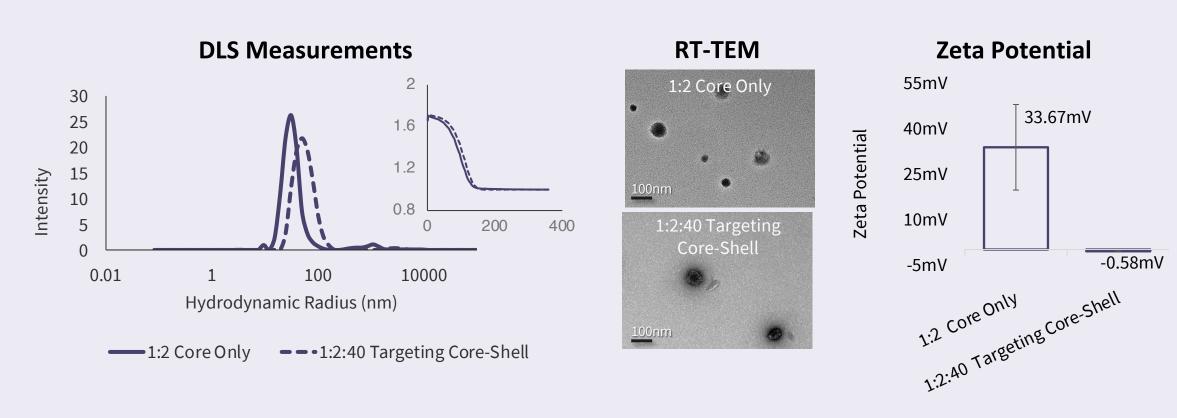
CCK-8 viability assay in murine lung fibroblasts was performed at increasing plasmid doses (0.5, 1, 2 µg/mL) and indicated retained viability in most groups.



RT-qPCR Analysis of TXNDC5 mRNA reveals significant knockdown in the targeted core-shell group.

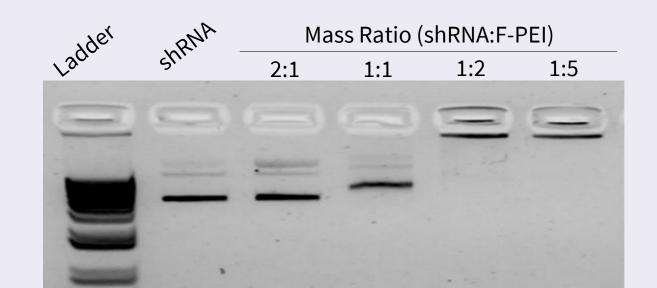
Size & Morphology Characterization of Micelle Nanostructures

F-PEI/shRNA Micelle



Size and Morphology Characterization

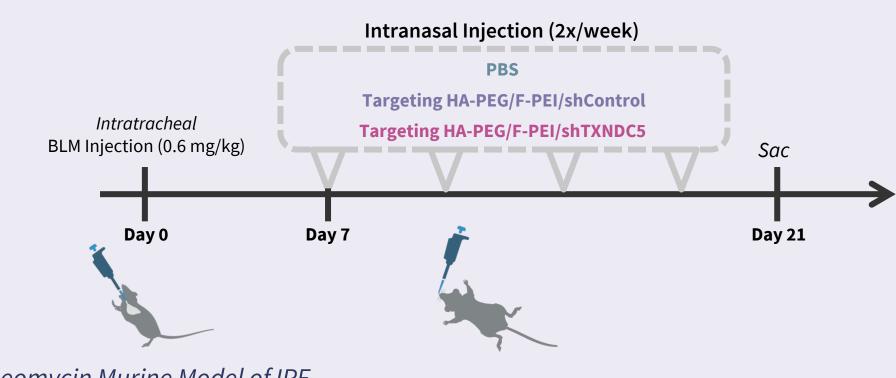
DLS reveals hydrodynamic radius of ~30 nm for cores and ~50 nm after shelling; TEM confirms uniform spherical morphology; zeta potential shifts from ~+34 mV (core) to ~-0.6 mV (core-shell).



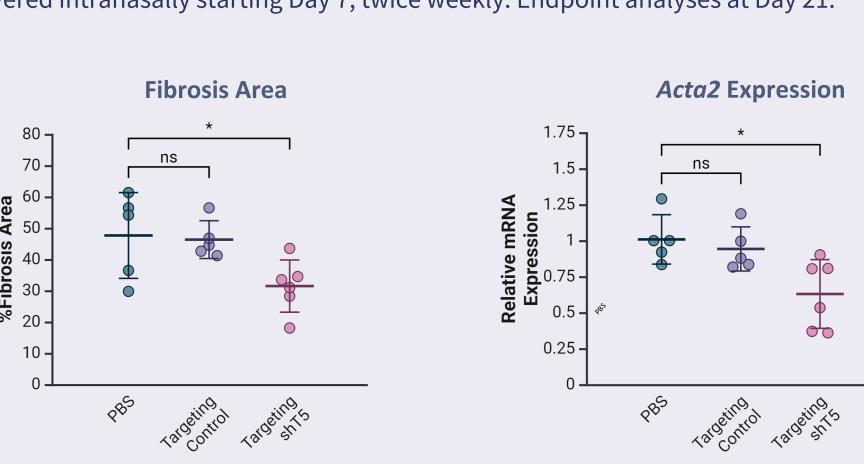
Retention Assay
Self-assembly with F-PEI enables
efficient plasmid condensation.
Gel electrophoresis confirms
complete shRNA complexation at
a 1:2 shRNA to F-PEI ratio.

In Vitro Experiments

In Vivo Studies Indicate Effective Reduced Fibrotic Burden & Downregulated Genetic Markers of Fibrosis

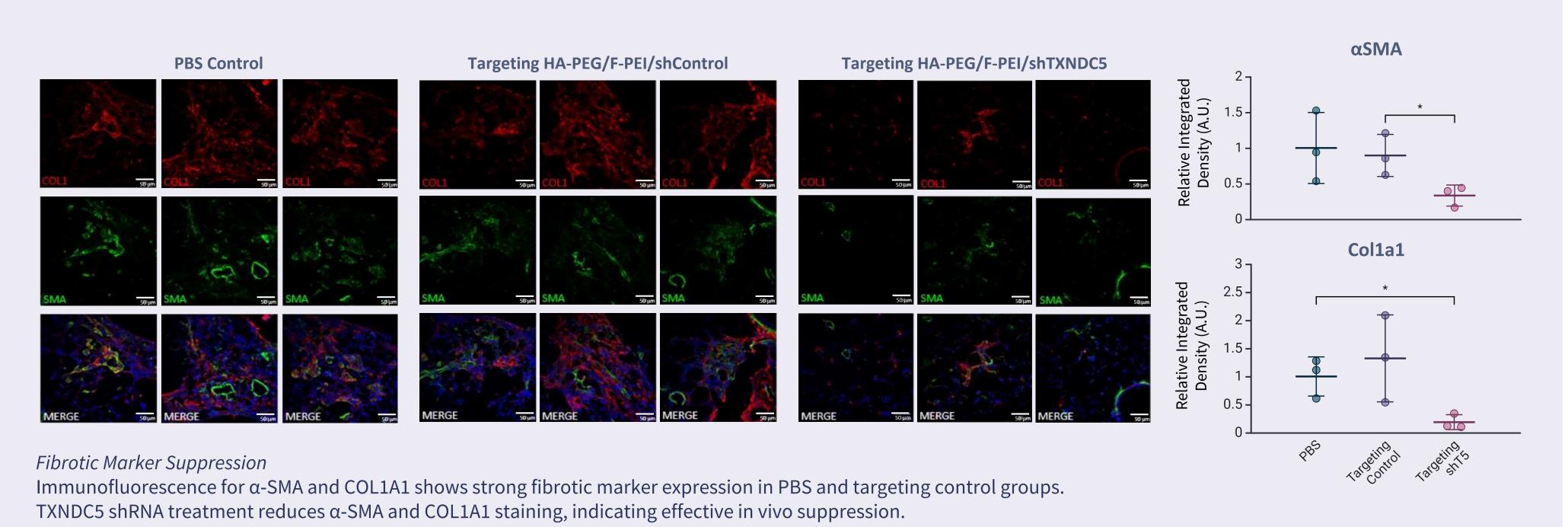


Bleomycin Murine Model of IPF C57BL/6 mice receive intratracheal bleomycin on Day 0 to induce fibrosis. Micelles are delivered intranasally starting Day 7, twice weekly. Endpoint analyses at Day 21.

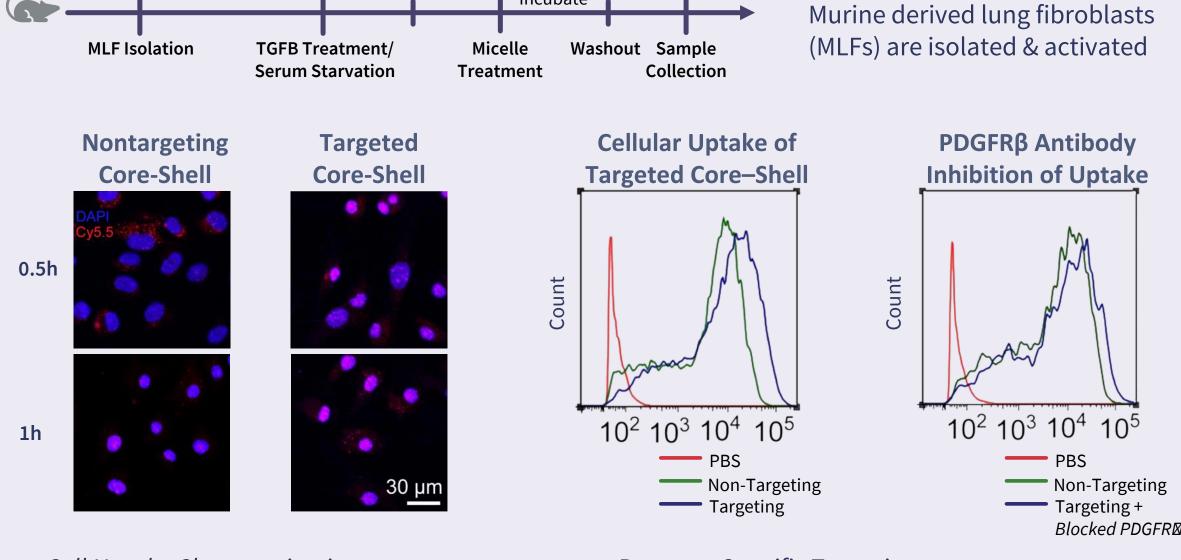


PBS Control F-PEI/shControl F-PEI/shTXNDC5 PBS Control F-PEI/shTXNDC5 PBS Control F-PEI/shTXNDC5

Lung Histology
Masson's Trichrome staining and relative Acta2 mRNA expression show high fibrosis in PBS and core- shell targeting control (carrying non-therapeutic plasmid) groups, indicating that the delivery vehicle alone does not alter fibrosis .TXNDC5 shRNA treatment reduces collagen deposition and Acta2 levels, confirming antifibrotic efficacy.



Efficient Uptake & Receptor-Specific Targeting In Vitro



Cell Uptake Characterization
Confocal microscopy shows Cy5-labeled
(red) micelles in fibroblasts at 0.5 and 1 h
post-treatment. Nuclei stained DAPI (blue).

Receptor Specific Targeting
Flow cytometry shows higher uptake of the targeted formulation with inhibition after presaturation using free anti-PDGFRβ antibody.

Future Perspectives

Future work will evaluate aerosolized delivery of the targeted core-shell micelles via nebulization, assessing particle stability, lung deposition, and biodistribution in healthy and fibrotic models. Additional studies will examine long-term lung function, immune response, and off-target effects to support safety and translation. The modular platform can be adapted for other RNA species, CRISPR systems, or small molecules, offering broad applicability for precision therapies.

References

Li, L., et al. (2017). ACS Nano, 11(1), 95–111. Lee, T., et al. (2020). Nature Communications, 11(1), 4254. Beljaars, L., et al. (2003). Biochemical Pharmacology, 66(7), 1307–1317. Lv, J., et al. (2014). Journal of Materials Chemistry B, 3(4), 642–650.

Acknowledgments

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