

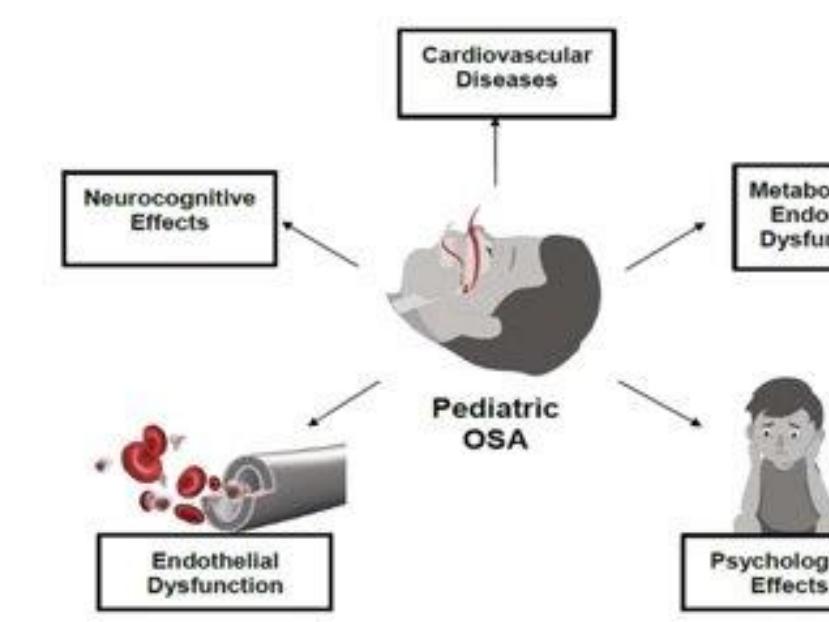


# Pediatric Obstructive Sleep Apnea is Associated with Localized Loss of Immune Tolerance in the Tonsils

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## Introduction

- Obstruction of the pediatric airway is in part thought to be the result of tonsillar hypertrophy
- Although there is a strong link between tonsillar hypertrophy and pediatric OSA, the pathogenesis of tonsillar hypertrophy in children is poorly understood
- Tonsillar hypertrophy may be related to a process of dysregulated immune cell signaling and inflammation.

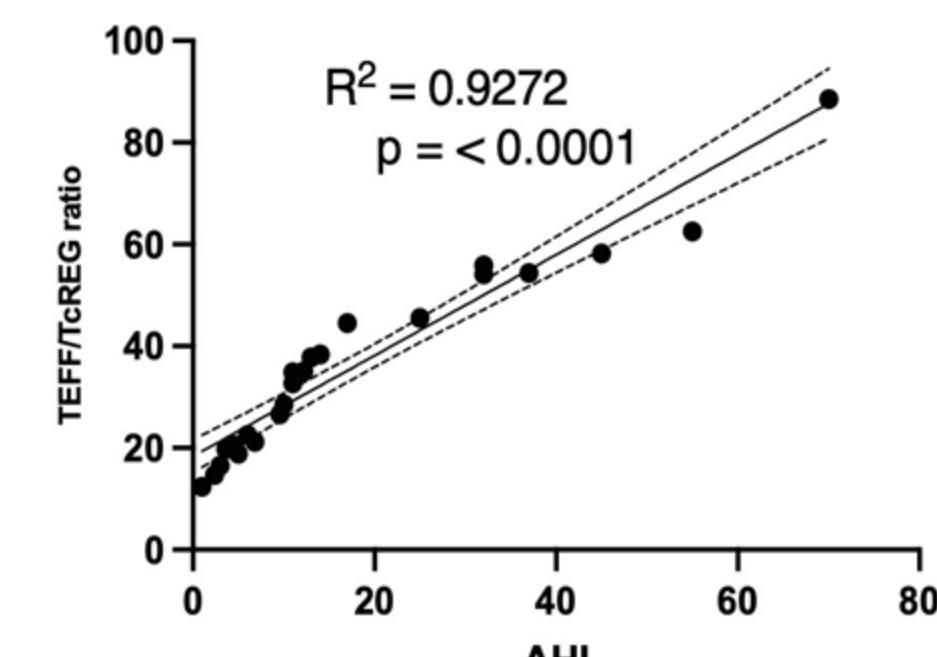


## Hypothesis

Loss of immune tolerance in the pediatric tonsil leads to decreased downregulation and an increased proinflammatory state, ultimately causing tonsillar hypertrophy and worsened OSA.

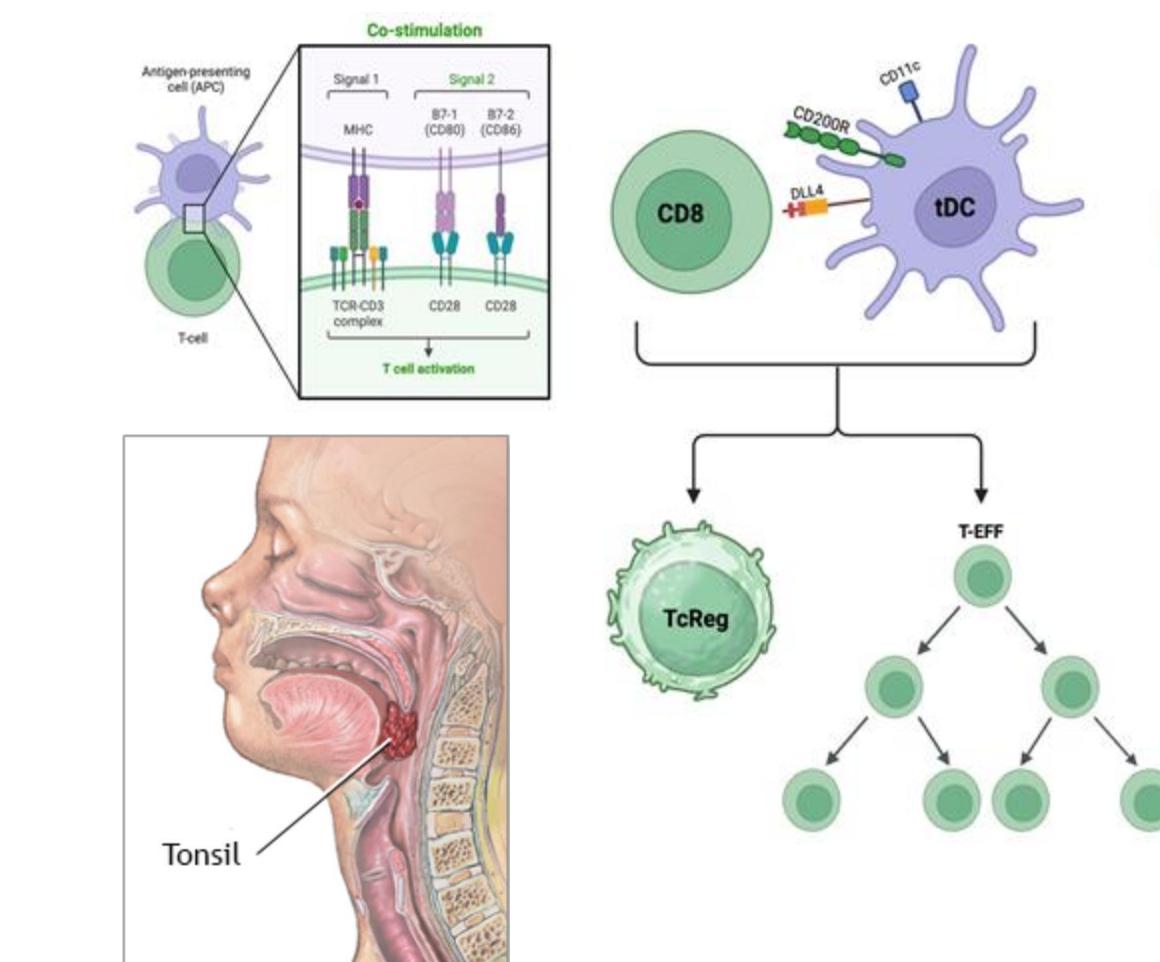
### Background- TEFF & TcREG

There are increased levels of TEFF cells (CD45+ CD3+ CD45RO+) and decreased levels of TcREG by flow cytometry with increasing AHI.



### Mechanism of Interest

- Interaction between tDCs and CD8 T cells leading to induction of T-regulatory cells and T-effector cells.
- Key elements of investigation involved analysis of dendritic cell markers CD11c, as well as CD200 glycoprotein receptor and DLL4 notch ligand.



## Methods

Analyzed three cohorts of patients for initial experiments  
AHI and BMI were controlled in all patients

Cohort #	n	Indication
Cohort 1	16	OSA
Cohort 2	10	Recurrent Tonsillitis
Cohort 3	6	Non-OSA

**Background** – Flow cytometry: TEFF & TcREG cells in patients with low vs high AHI.

**Figure 1** – Flow cytometry: Dendritic cells in patients with low vs high AHI.

**Figure 2** – Co-culturing: tDC induction of TcREG: 4 days of co-culturing tDCs with native CD8 T cells → Anti-CD3 antibody crosslinked with TCR → Induction of TcREG measured in patients with OSA vs no OSA.

**Figure 3** – Single Cell RNA Sequencing comparing cell types in low vs high AHI.

Two additional patients were collected for staining  
BMI was controlled in both patients

Patient #	AHI	BMI
Patient 1	3.9	17.3
Patient 2	39	16.5

**T-cells – Immunohistochemistry Staining:** Comparison of the % Area of CD3+ T-cells between low and high AHI. Comparison of the number of FoxP3+ cells/15x mag field.

**T+DC – Immunohistochemistry Staining:** Comparison of the % Area of overlap between CD3+ T-cells and CD11c+ dendritic cells between low and high AHI.

**Dendritic Cells (DC) – Immunohistochemistry Staining:** Comparison of the % Area of CD11c+ dendritic cells between low and high AHI.

### Immunohistochemical staining – TcREG & Teff Cells

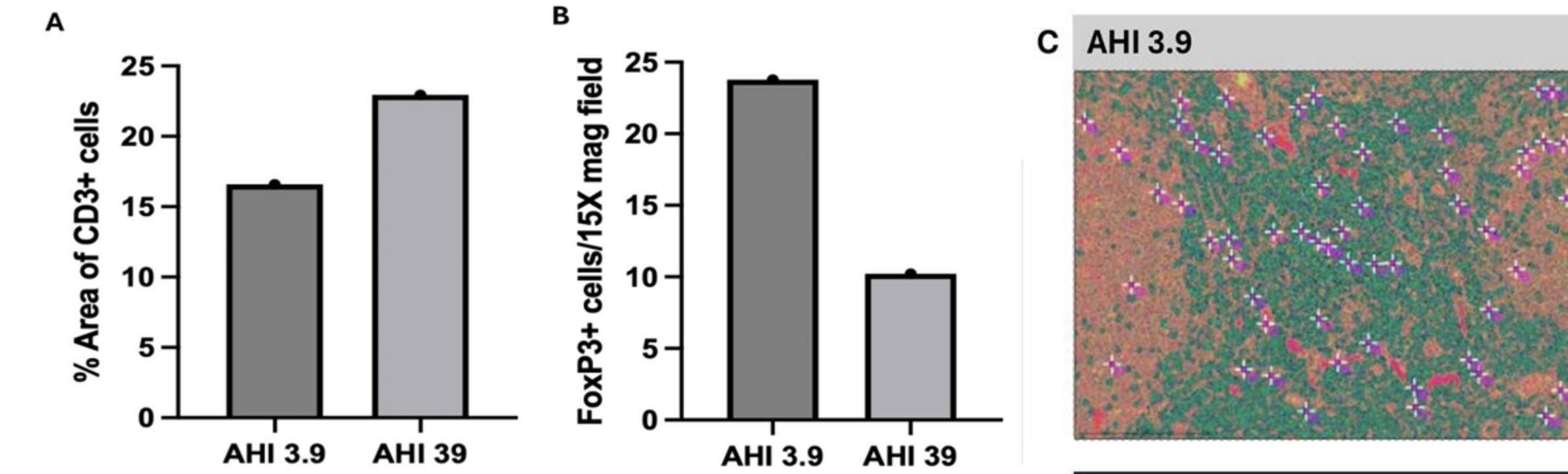
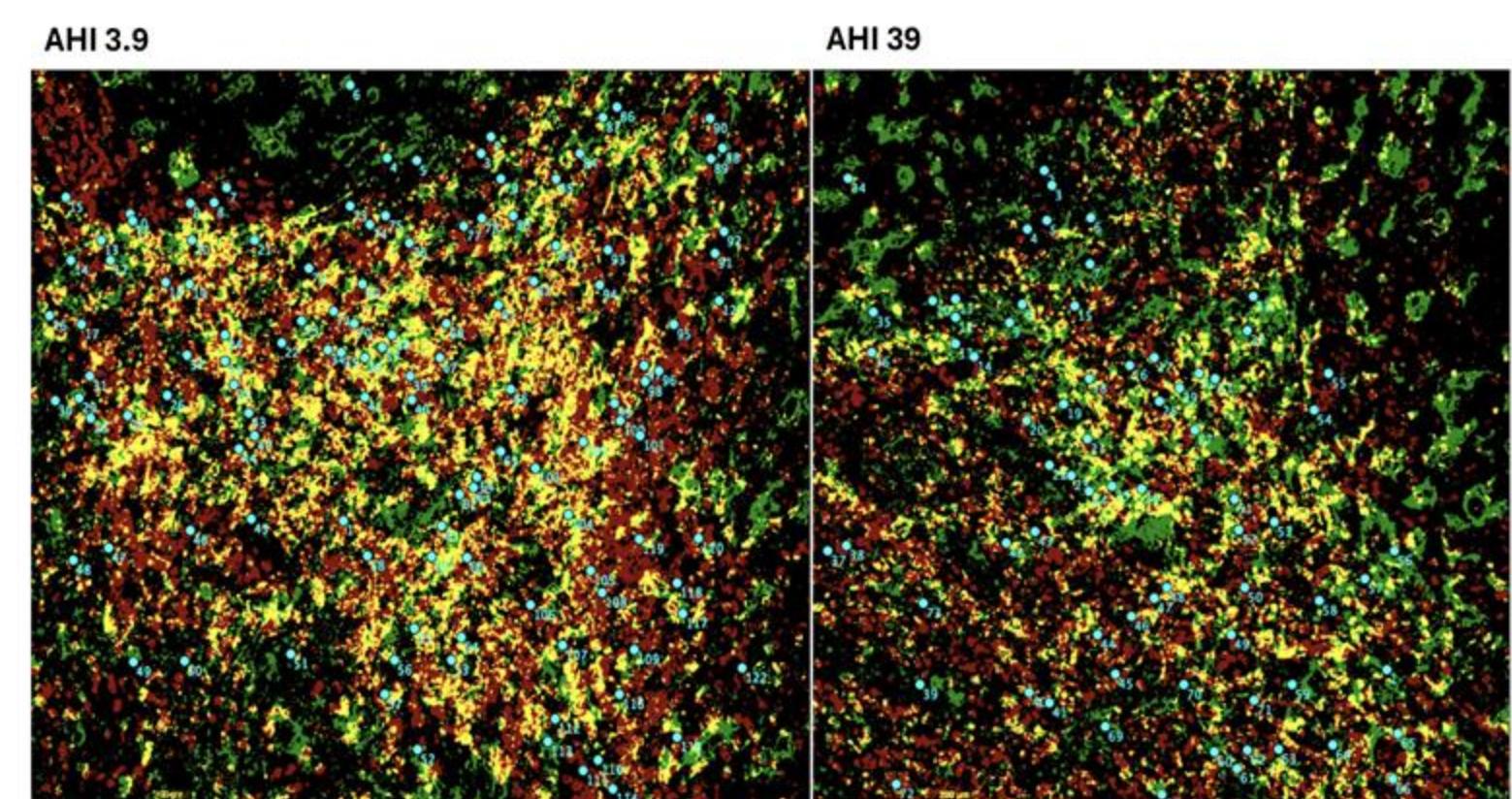


Figure A represents % area of CD3+ cells between low and high AHI patients. Figure B represents the number of FoxP3+ cells between low and high AHI patients. Images C and D are visual representations of CD3+ cells (green) with FoxP3 TcREGs overlaid (magenta dot with white crosshairs)



### Immunohistochemical staining – T-cells and DC Overlap

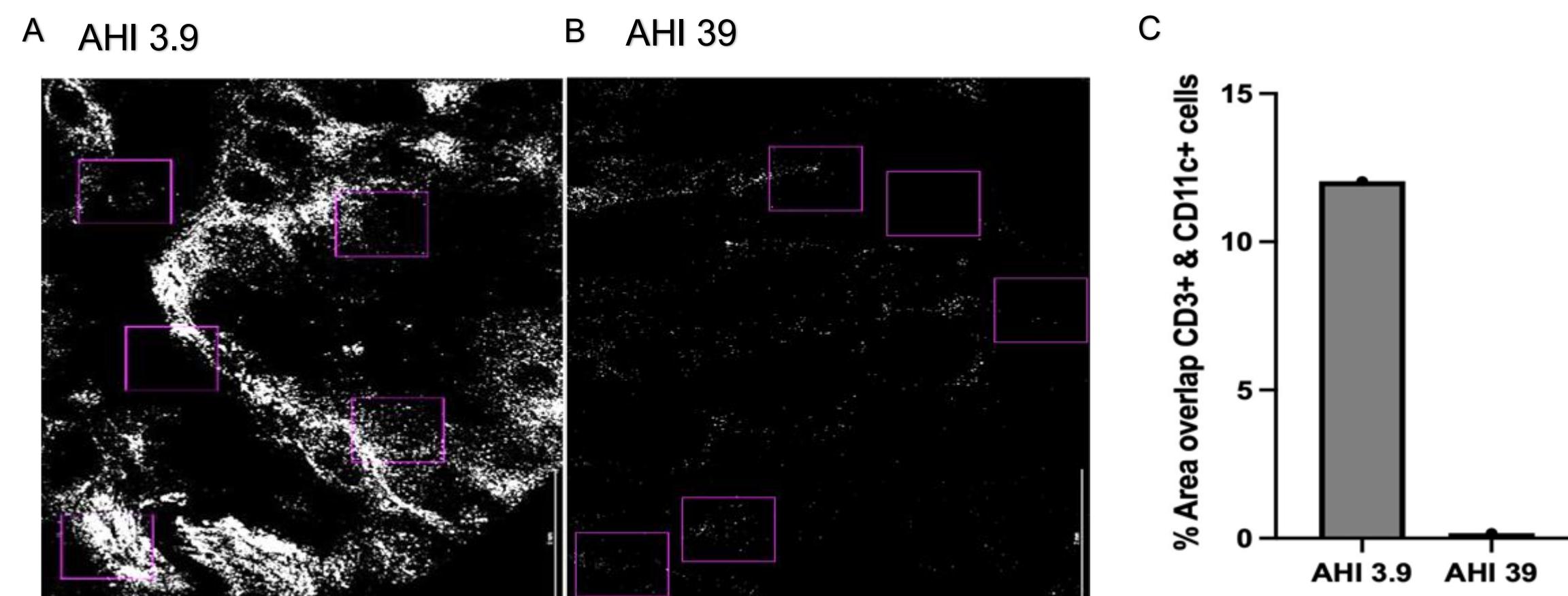


Image A shows colocalization between CD3+ T-cells and CD11c+ tDC for the entire tonsillar cross section. Figure B is a graphical representation of the % area overlap shown in Image B.

### Immunohistochemical staining – Dendritic Cells

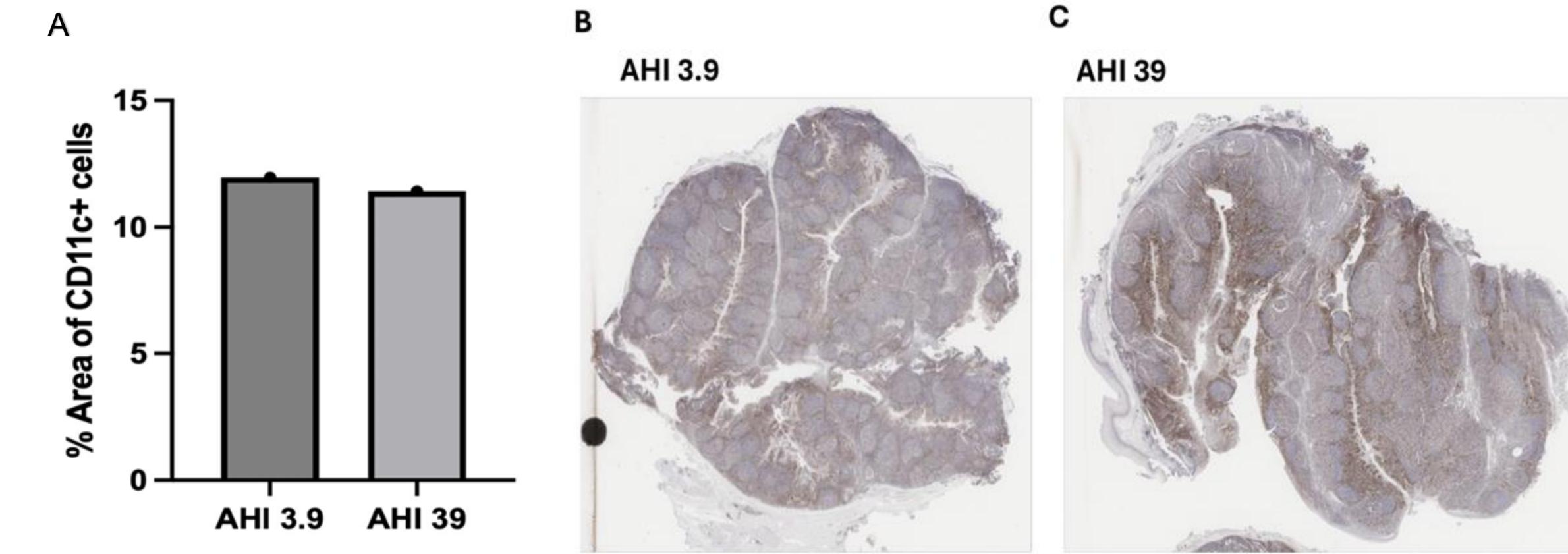
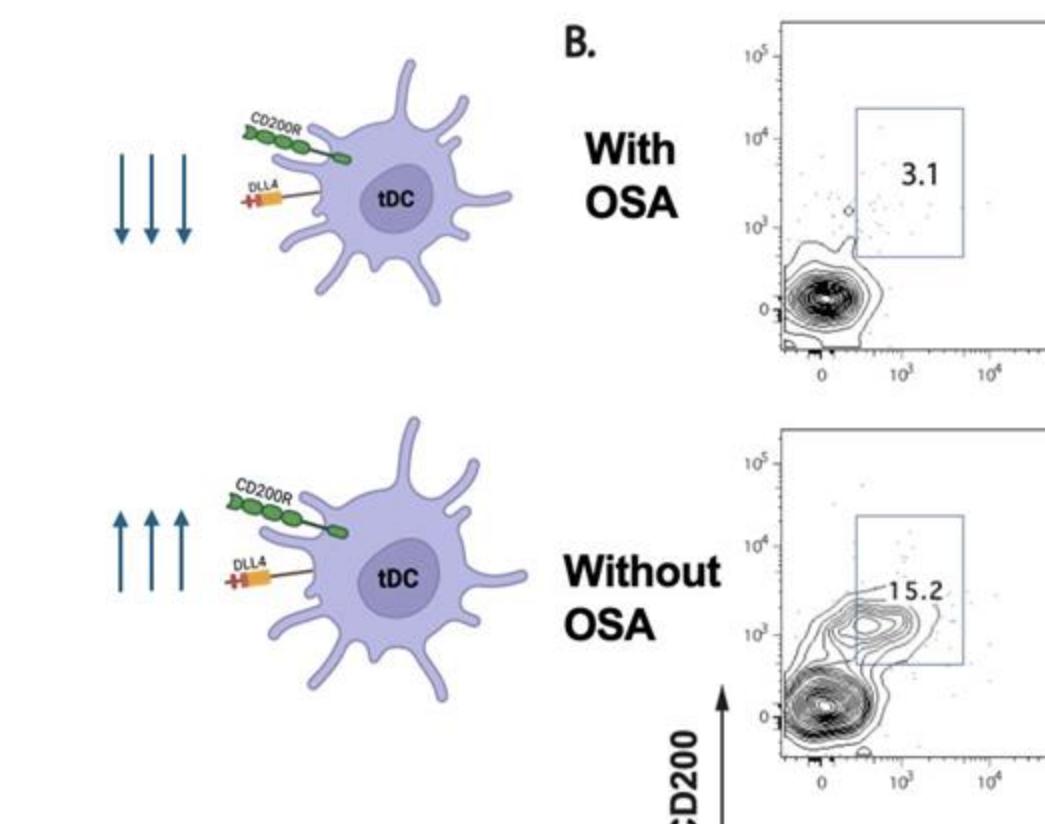


Figure A represents flow cytometry and shows no difference in CD11c+ tDCs between low and high AHI. Images B and C are gross images of each patient's tonsils stained with CD11c. Importantly, CD11c does not differentiate between DLL4+ or DLL4- dendritic cells. Dendritic cells localize to different locations in tonsils from patients with higher AHI.

## Results

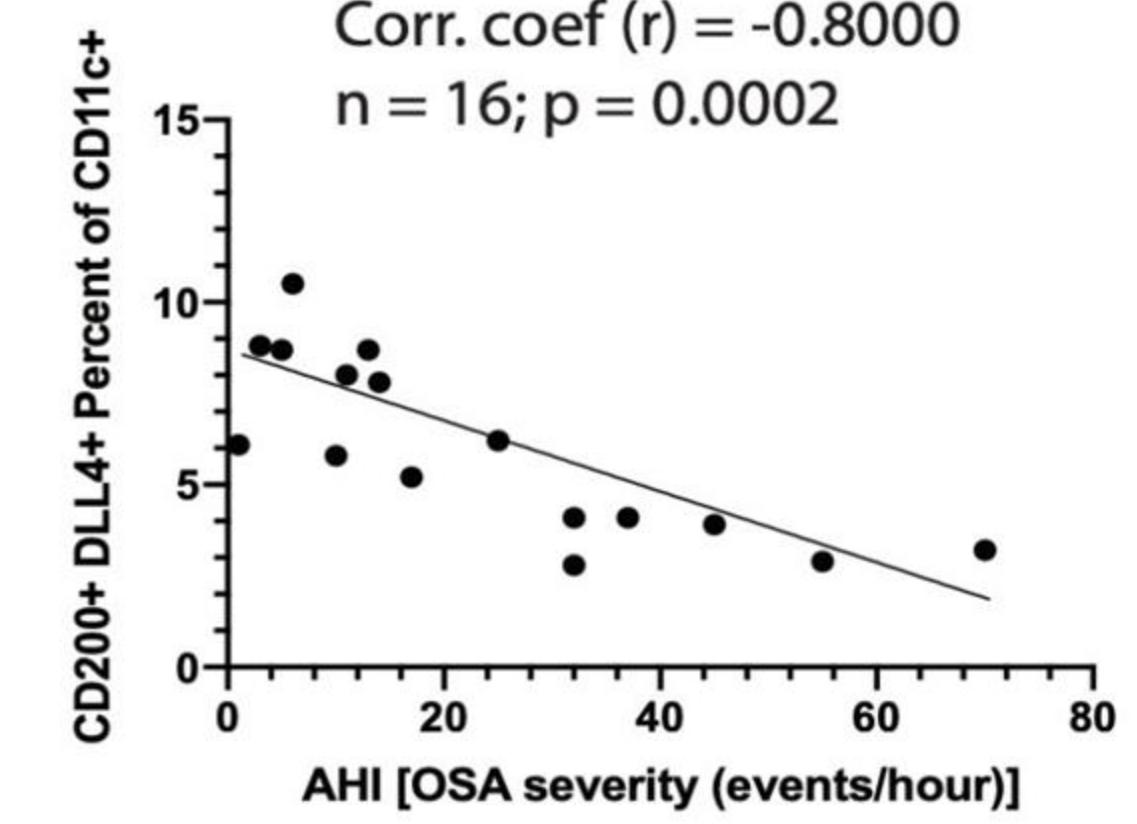
### Figure 1 - Flow Cytometry - Dendritic Cells

There were increased tDC (CD200+ and Notch ligand DLL4+) - APCs known to induce TcREG (FoxP3) and CD25 in CD8 T-cells in patients without OSA



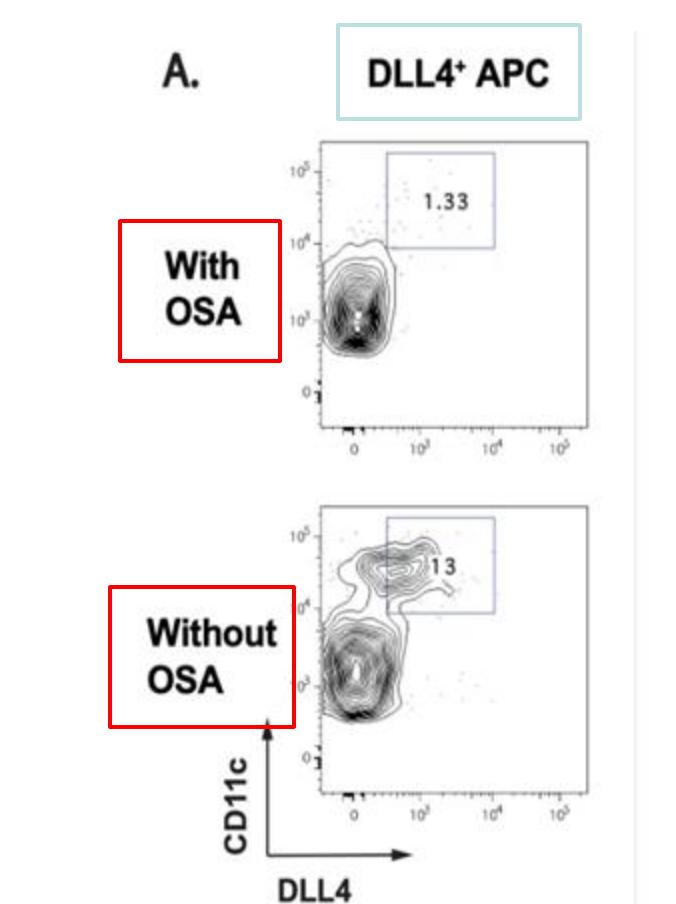
Levels of CD200 DLL4 double-positive subset of DC (tDC) negatively correlated with AHI

Corr. coef (r) = -0.8000  
n = 16; p = 0.0002

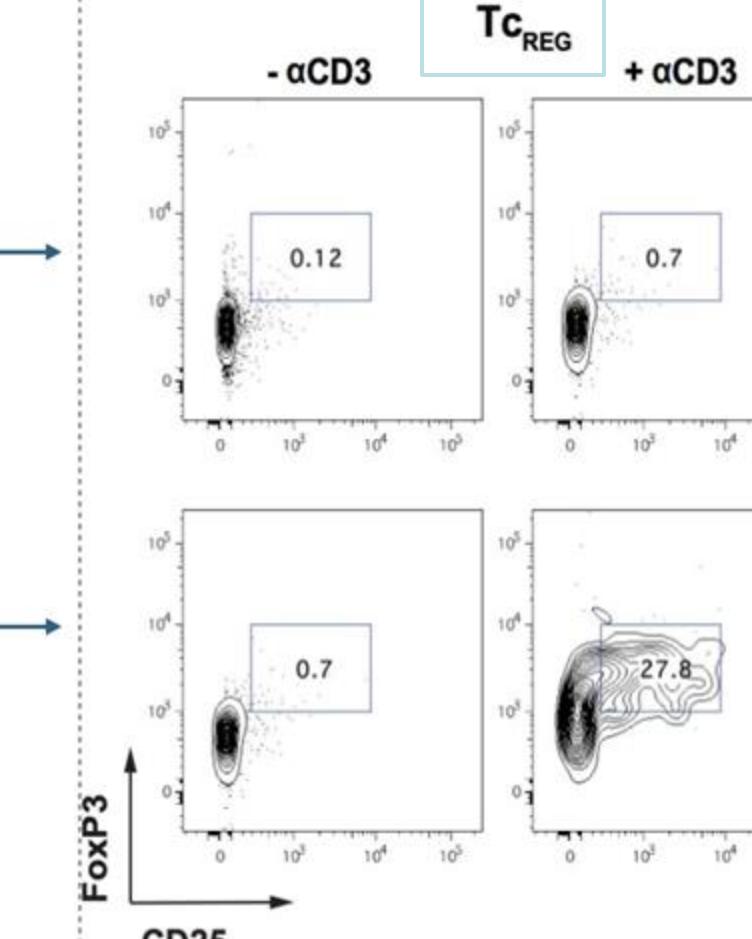


### Figure 2 – tDCs induce TcREG

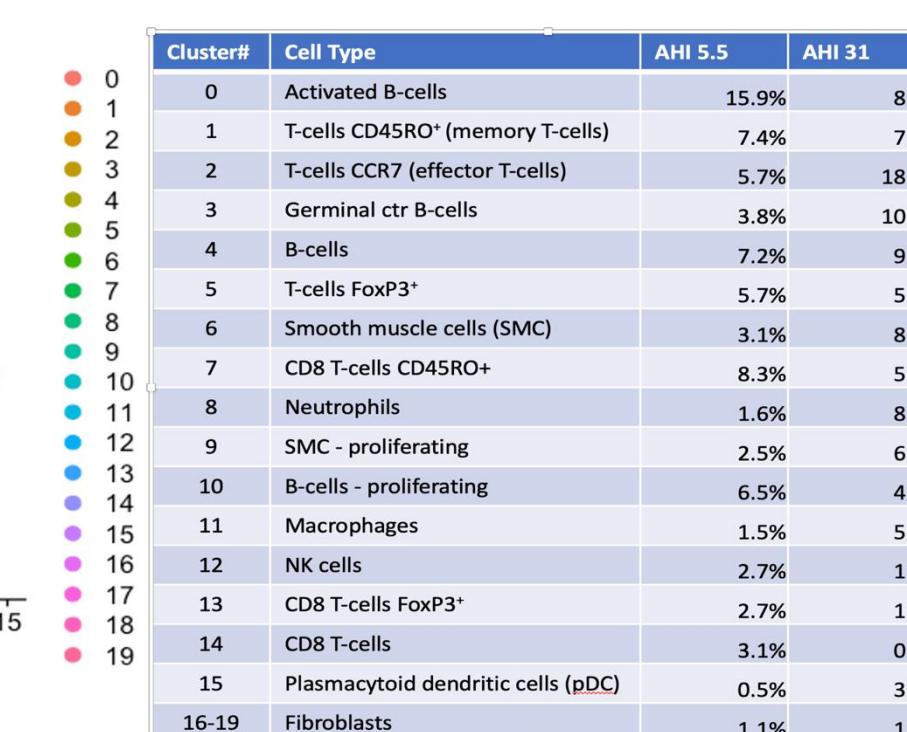
tDCs are more robust from patients with tonsillitis vs those with OSA.



These tDCs were more abundant after 4 days of co-culturing and ultimately led to higher numbers of TcREG induction.



Relationship between levels of TcREG induction and DLL4+ DC expression



## Conclusion

The results of flow cytometry and scRNA-seq indicate there is significant expansion of effector T-cells (cluster 2), germinal center B-cells (cluster 3), and smooth muscle cells (cluster 9). There were decreased but not statistically significant levels of FoxP3+ T-cells in patients with higher AHI.

Although we observe a loss of tDCs with increased AHI, we have not yet identified the mechanism promoting increased T-EFF cells. The Teff may be activated by an alternative APC pathway eg. B cells, leading to proinflammatory T-cell expansion. This represents an avenue of future study.

