TuNa-Al: A Hybrid Kernel Machine to Design Tunable Nanoparticles for Drug Delivery







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Technology Exposition

- Drug-excipient nanoparticles are an emerging drug delivery platform.
- They are known for facile synthesis through self-assembly, high drug-loading capacity, and a rational design process informed by machine learning^{1,2}.
- However, their simple synthesis (Fig. 1) also prevents tuning of material composition.

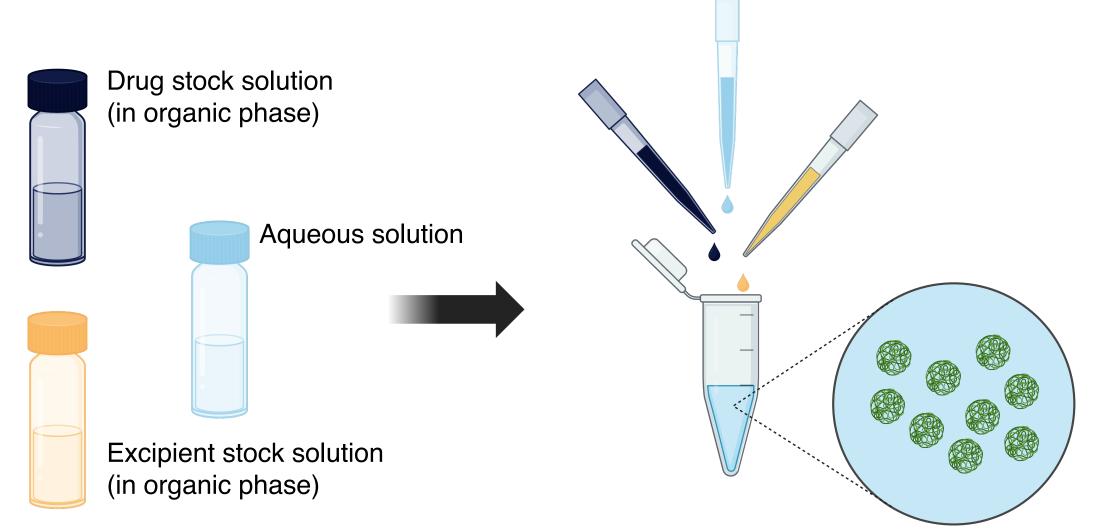


Figure 1. Schematic of the drug-excipient nanoparticle synthesis protocol. The an organic solvent (e.g., DMSO) and undergoes phase reversal upon the addition of an aqueous solution, leading to the self-assembly of nanoparticles.

Aim

- Devise a robotic-assisted synthesis protocol to create nanodrugs at various excipient/drug molar ratios.
- Design a **hybrid kernel machine** approach to prediction nanoparticle formation.
- Apply the computational model to identify novel drug-excipient nanoparticles.

Kernel Hybridization

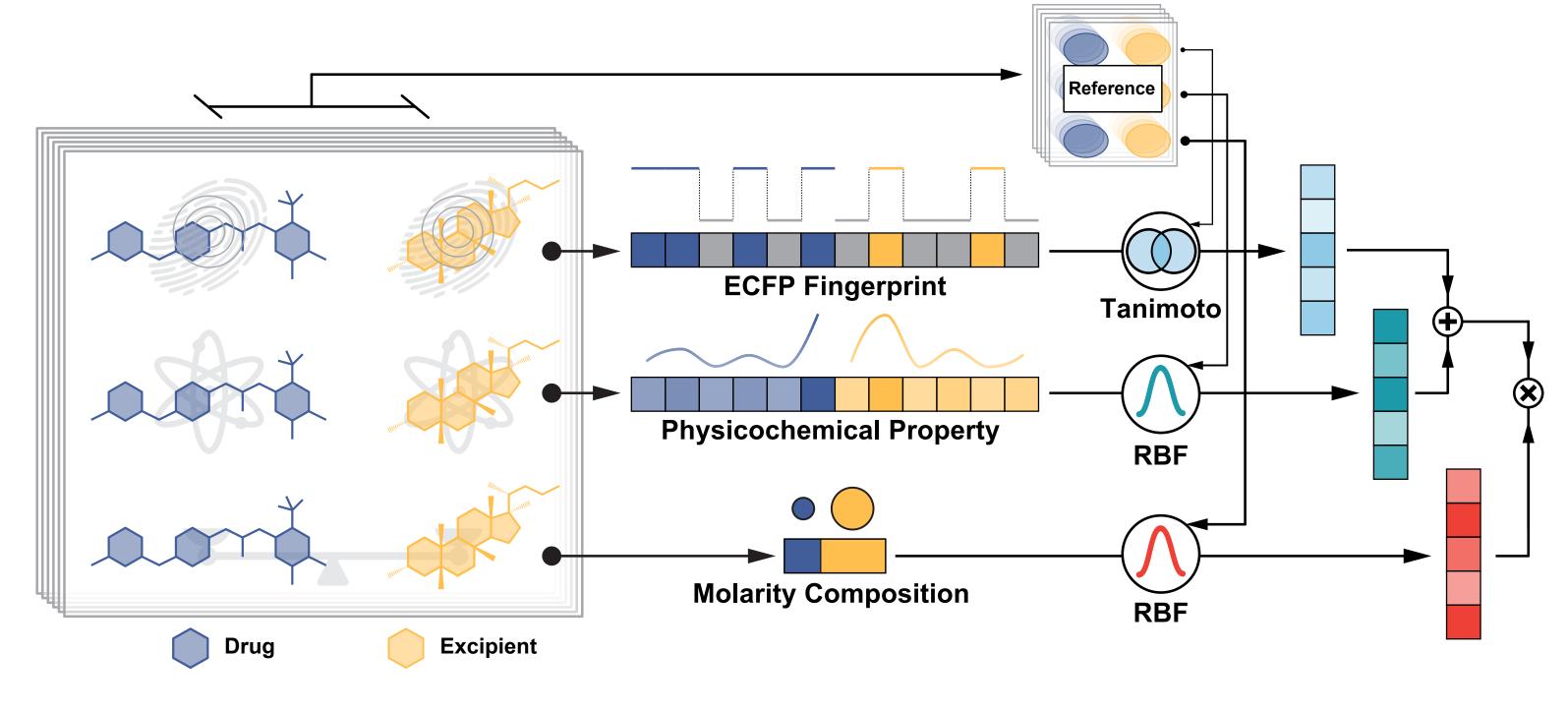


Figure 3. Schematic of the hybrid kernel machine design. Hybrid kernel machine architecture, featuring inputs from binary ECFP fingerprints, continuous physicochemical properties, and molarity ratios of drug and excipient during synthesis. RBF, radial basis function.

Automated Exploration

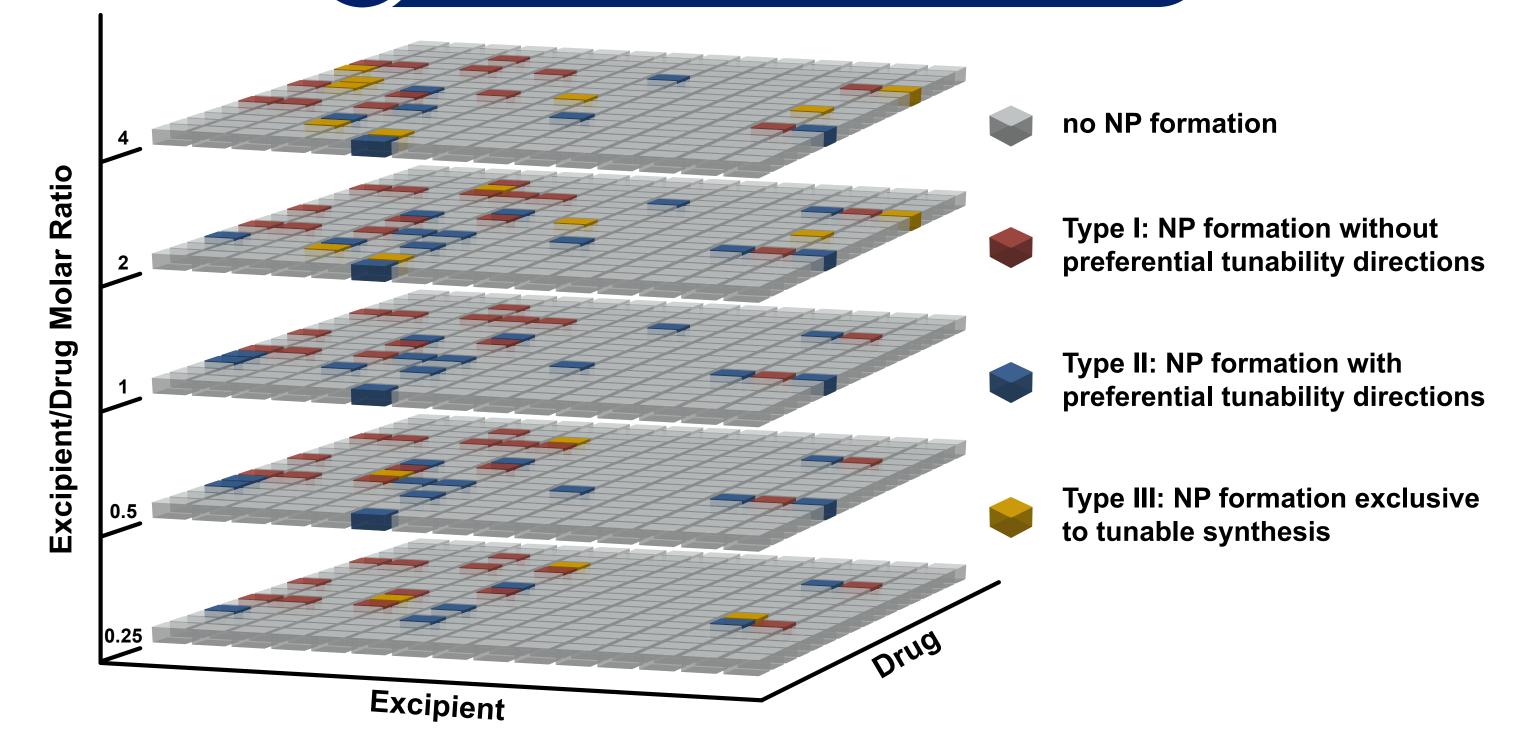
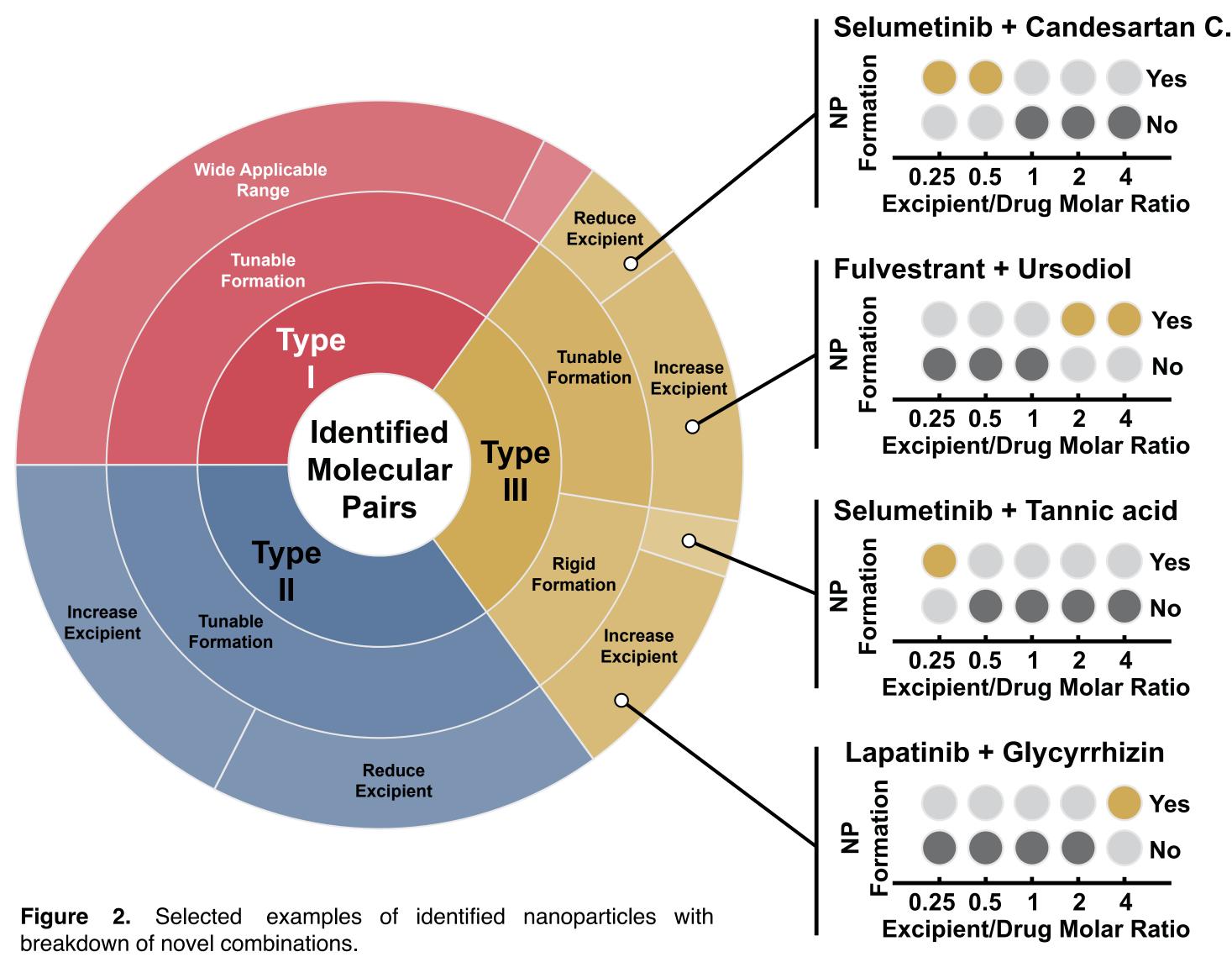


Figure 2. Robot-assistant screening results of nanoparticle formation experiments. The full data matrix includes 17 drugs and 15 excipients. The optimized synthesis protocol investigated five different excipient/drug molar ratios (0.25, 0.5, 1, 2 and 4) in order to expand the nanoparticle searching space.



Computational Evaluation default kernel 1.0 — 1.0 hybrid kernel **** score **ROC-AUC** Normalized 0.6 other models in comparison **False Positive Rate**

Figure 4. Evaluation of the hybrid kernel machine on predicting nanoparticle formation. a, Evaluation of kernel or our new hybrid kernel. Models with default kernels (SVM and GP with RBF kernel; kNN with Minkowski distance) are shown in gray, while hybrid kernel models are highlighted in blue. b,c, ROC curves and AUC scores of all surveyed models. The best-performing support vector machine (SVM) model is shown in red, while all other models are shown in shades of gray. The dashed diagonal represents random guessing (ROC-AUC = 0.5), and the shaded areas indicate one standard deviation across five independent cross-validation results for each model. SVM, support vector machine; GP, Gaussian process; RF, random forest; MPNN, message-passing neural network; kNN, k-nearest neighbors; MLP, multi-layer perceptron. d, Computational cost comparison (normalized to SVM CPU time, except the MPNN which uses GPU acceleration). Unpaired t-test ($\alpha = 0.05$); ****p < 0.0001.

Congo red (CR)

Standard NP

(100% CR)

Tunable NP

(25% CR)

Standard NP Tunable NP

(100% CR) (25% CR)

Nanodrug Innovation increasing excipients – formulate hard-to-load drug **Molar Ratio Prediction Validation** 40 nm Venetoclax 60 nm >1000 nm >1000 nm 0.5 0.25 >1000 nm 0.25 positive prediction Taurocholic acid (TCA) **Excipient/Drug Molar Ratio** negative prediction Venetoclax + TCA 1000 + TCA 100 Venetoclax 10000 Radius (nm) g 🕏 120 ┐ iability -08 -08 80 -**60** -50 -Venetoclax Venetoclax + TCA + TCA

Figure 5. Venetoclax-Taurocholic Acid (TCA) nanoparticles form when TCA is added more than Venetoclax. a, Chemical structures of venetoclax and taurocholic acid (TCA). b,c, Model predictions and experimental validation (hydrodynamic radius) of venetoclax-TCA nanoparticles at different molar ratios. **d-f**, Transmission electron microscope (TEM) images, size distribution, and dispersion stability of 500 μ M venetoclax, both unformulated and TCA-formulated (venetoclax:TCA = 2, molar ratio). **g**, Venetoclax nanoparticles exhibit improved cytotoxicity over free drugs on Kasumi-1 acute myeloblastic leukemia (AML) cells.

Research Supported by

Time (h)

BIOMEDICAL

ENGINEERING

National Institute of

Venetoclax Concentration (µM)

Affiliated Institution 1. Department of Biomedical Engineering, Duke University, Durham, NC 27708

2. Department of Medicine, Duke University School of Medicine, Durham, NC 27710

0.25 0.5

Excipient/Drug Molar Ratio

Formulation Injection

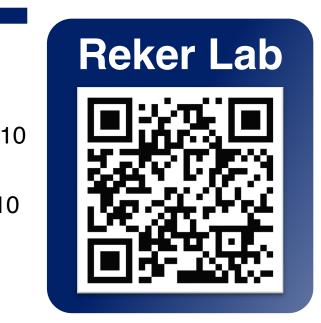
10000

3. Pharmacokinetics/Pharmacodynamics (PK/PD) Core Laboratory, Duke Cancer Institute, Durham, NC 27710 4. Department of Radiation Oncology, Duke University School of Medicine, Durham, NC 27710

10 min

- 5. Department of Pharmacology and Cancer Biology, Duke University School of Medicine, Durham, NC 27710 6. Department of Surgery, Pathology and Integrative Immunobiology, Duke University, Durham, NC 27708
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General Medical Sciences



Standard NP (100% CR)

reducing excipients – maintain bioequivalence

Trametinib

18 nm

25 nm

50 nm

55 nm

62 nm

0.25

Sample Collection

(10 min, 30 min,

1.5 h, 6 h, 24 h)

Post-injection Time (h)

drug concentration profiles show largely bioequivalent behavior. Unpaired t- test ($\alpha = 0.05$); n.s., $p \ge 0.05$, *p < 0.05, *p < 0.01, ***p < 0.001, ****p < 0.001, ****p < 0.0001.

Figure 6. Synthesizing bio-equivalent trametinib nanoparticles using less Congo red (CR). a, Chemical structures of trametinib and Congo red (CR). b,c, Model

predictions and experimental validation of Trametinib-CR nanoparticles. d, TEM images of 500 µM trametinib. e, Drug loading of trametinib nanoparticles at CR/trametinib

molar ratios of 1:1 (standard nanoparticles, 100% CR) and 1:4 (tunable nanoparticles, 25% CR). f, Standard and tunable trametinib nanoparticles (20 μΜ) exhibit

comparable in vitro cytotoxicity against HepG₂ human liver cancer cells. g, Schematic of in vivo experiment. h, Plasma drug concentration following retro-orbital injection of

equal doses of standard and tunable trametinib nanoparticles. i, Key pharmacokinetic parameters of standard and optimized trametinib nanoparticles derived from plasma

positive prediction

Drug Quantification

PK Analysis

■ Standard NP (100% CR)

Tunable NP (25% CR)

Preprint

Tunable NP (25% CR)

+ CR

를 200

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